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16 Running title:
17 Immune abilities of pcDNA3 and pSINCP encoding FMDV
18 P1-2A3C3D
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21 **Daniel Dory^a, Michelle Rémond^b, Véronique Béven^a, Roland Cariolet^c, Stephan**
22 **Zientara^b and André Jestin^a**
23

24 ^a Viral Genetics and Biosafety Unit, French Food Safety Agency (Afssa), Fr-22440,
25 Ploufragan, France

26 ^b UMR 1161 (Inra-Afssa-Envva), Afssa, Fr-94703, Maisons-Alfort, France

27 ^c Healthy Pigs Production and Experimentation Section, Afssa, Fr-22440, Ploufragan, France
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38 *Corresponding author: Daniel Dory, Viral Genetics and Biosafety Unit, Afssa (French Food
39 Safety Agency), BP-53, Fr-22440, Ploufragan, France
40 Phone: 00 33 2 96 01 64 42
41 Fax: 00 33 2 96 01 62 83
42 d.dory@afssa.fr
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Foot and Mouth Disease Virus neutralizing antibodies production induced by pcDNA3 and Sindbis virus based plasmid encoding FMDV P1-2A3C3D in swine

Abstract

DNA vaccination against FMDV is an attractive and alternative strategy to the use of classical inactivated viral vaccines. The injection of a pcDNA3.1-based DNA vaccine encoding for FMDV P1-2A3C3D and GM-CSF proteins had previously been shown to induce the production of neutralizing antibodies against FMDV and partially protect swine against an experimental challenge. Based on the induction of FMDV humoral immune responses, the aim of the present study was to see if the Sindbis virus derived plasmid (pSINCP) backbone could advantageously replace the pcDNA3.1 one in DNA immunization against FMDV in swine. For this purpose, groups of 3 or 4 pigs received three injections by intramuscular route, intradermal route or an association of both routes, at 2 to 3 week intervals. The pcDNA3.1 based DNA vaccine was shown to induce the production of higher amounts of FMDV neutralizing antibodies after intradermal injection. Intramuscular injection of the same vaccine, or intramuscular and/or intradermal injection of the pSINCP-based DNA vaccine resulted in a significantly lower induction of FMDV neutralizing antibodies. In conclusion, the humoral immune response of a DNA vaccine encoding for FMDV P1-2A3C3D was not improved by the pSINCP backbone and was higher when the plasmids were injected by the intradermal route.

67 **1. Introduction**

68

69 Foot-and-Mouth Disease Virus (FMDV) is the aetiological agent of an important
70 disease of livestock. FMD is highly contagious and affects cloven-hoofed animals, mostly
71 cattle, swine, sheep and goats. FMDV belongs to the Aphthovirus genus of the Picornaviridae
72 family (Rodrigo and Dopazo, 1995, Sobrino et al., 2001). The positive-strand RNA genome
73 of about 8500 nucleotides length is enclosed within a protein capsid. The viral open reading
74 frame (ORF) encodes a single polyprotein that is cleaved by viral proteases to yield different
75 structural and non-structural proteins (Ryan et al., 1989).

76 Regular vaccination is one of the strategies employed to control disease propagation,
77 and has resulted in eradication of the disease in some parts of the world (particularly Western
78 Europe) (Sobrino et al., 2001). Efficient vaccination was achieved with adjuvanted chemically
79 inactivated FMD virus, which induced a consistent humoral response. Nevertheless, this kind
80 of vaccination does have some disadvantages: a) the production of FMDV requires high
81 containment facilities and the risk of virus escape is still possible (Grubman and Baxt, 2004,
82 Record, 2007), b) inactivation of the virus may be incomplete in some cases (King et al.,
83 1981), c) the induced protection is short lasting (Cox et al., 2003) and d) discriminating
84 between vaccinated and infected animals can be a problem with some vaccines due to the lack
85 of validated differentiation techniques. For all these reasons, alternative vaccination strategies
86 such as the use of proteins, peptides, replicating vectors, attenuated strains and DNA vaccines
87 have been investigated (Grubman and Baxt, 2004, Sobrino et al., 2001). Among these, DNA
88 vaccination presents several advantages. In fact, the production of DNA vaccines is easy and
89 safe, long-term storage of the vaccine is possible, constructs encoding fusion proteins can be
90 developed and DNA vaccines can serve as marker vaccines. This technology can also be used
91 efficiently to create vaccines against emerging serotypes. Numerous trials have been carried

92 out in this context to obtain an efficient DNA vaccine against FMDV, with various degrees of
93 success. Plasmids encoding FMDV VP1 (Park et al., 2006, Xiao et al., 2007) or FMDV
94 epitopes (Cedillo-Barron et al., 2003, Chen and Shao, 2006, Du et al., 2007, Zhang et al.,
95 2003) were tested. Three injections of a plasmid encoding the viral structural protein
96 precursor P1-2A and the non-structural proteins 3C and 3D, together with a plasmid encoding
97 GM-CSF, induced the production of FMDV-specific and neutralizing antibodies and partially
98 protected pigs from an experimental FMDV infection (Cedillo-Barron et al., 2001). It is
99 apparent from these results that the efficiency of this DNA vaccine needs to be considerably
100 improved. In fact, the number of injections required is too high for a vaccine that must be able
101 to act rapidly in the case of a FMDV outbreak. Stronger FMDV specific and neutralizing
102 antibody responses were induced by increasing the amount of plasmids (Li et al., 2006), but
103 not by co-injecting a plasmid encoding the BAFF protein (Bergamin et al., 2007).
104 Interestingly, two co-injections of the FMDV and GM-CSF constructs, both formulated with
105 D,L-lactide-co-glycolide based particles, induced humoral and cytotoxic immune responses
106 and protected 5 out of 5 sheep against a virulent FMDV challenge (Niborski et al., 2006).

107 A new generation of non-replicating plasmids derived from the Sindbis virus
108 (pSINCP) was generated in 1996 (Dubensky et al., 1996). This approach involved the
109 conversion of a self-replicating vector RNA (replicon) into a layered DNA-based expression
110 system. The first layer includes a eukaryotic RNA polymerase II expression cassette that
111 initiates nuclear transcription of an RNA which corresponds to the Sindbis virus vector
112 replicon. After transport of this RNA from the nucleus to the cytoplasm, the second layer
113 proceeds according to the Sindbis virus replication cycle and results in expression of the
114 heterologous gene. For example, increased efficacies of anti-herpes simplex virus (Hariharan
115 et al., 1998) and anti-*Mycobacterium tuberculosis* (Kirman et al., 2003) vaccinations were
116 obtained with these plasmids. Furthermore, we demonstrated that a single injection of 13 µg

117 of pSINCP encoding Pseudorabies virus (PrV) glycoproteins gB, gC and gD, i.e. 25 times less
118 than for pcDNA3, efficiently protected pigs against a highly virulent experimental PrV
119 challenge (Dory et al., 2005).

120 The aim of the present study was to compare the level of immunization of pigs
121 injected with the FMDV P1-2A3C3D construct cloned either in pcDNA3.1 or in pSINCP.
122 Either pcDNA3.1/GM-CSF or pSINCP/GM-CSF was used as adjuvant in the corresponding
123 groups. Two quantities of FMDV construct-encoding plasmids were tested: the one originally
124 used by Cedillo-Barron (Cedillo-Barron et al., 2001) and one 25 times smaller, as used in our
125 previous PrV study (Dory et al., 2005). In a second part of the study, the efficacies of pig
126 immunization by intramuscular (IM) or intradermal (ID) injection were compared.

127

128 **2. Materials and methods**

129 *2.1. Plasmids*

130 pcDNA3.1 plasmid encoding the FMDV O₁K P1-2A3C3D sequence (Cedillo-Barron et al.,
131 2001), and 9829 bp in length, was kindly provided by Paul Barnett (IAH, Pirbright, UK). The
132 P1-2A3C3D cassette was extracted by a blunt *Pme* I digestion and inserted into the
133 dephosphorylated blunt *Pml* I site of the pSINCP plasmid (kindly provided by John Polo,
134 Chiron Corporation, USA) (Fig. 1). pSINCP encoding FMDV O₁K P1-2A3C3D sequences
135 was selected by endonuclease restrictions, PCR and sequencing (not shown). The resulting
136 plasmid was 16306 bp long. 4.5×10^5 porcine kidney-derived PK15 cells per well were
137 incubated for 24 hours at 37°C in a 6-well plate. These 70-80 % confluent cells were then
138 transfected either with 2 µg of pcDNA3.1/P1-2A3C3D, pSINCP/P1-2A3C3D, empty-
139 pcDNA3.1 or empty-pSINCP by using lipofectamine plus transfection reagent (Invitrogen,
140 Gaithersburg, MD, USA) according to the manufacturer's instructions. Forty-eight hours
141 later, the expression was determined by immunostaining with a mouse anti-VP1 monoclonal
142 antibody B2 (kindly provided by Emiliana Brocchi, IZS, Brescia, Italy) (Cedillo-Barron et al.,
143 2001) followed by incubation with a HRP-conjugated goat anti-mouse antibody and the
144 peroxidase AEC substrate (Serotec Ltd, Oxford, UK) (Fig. 1). pCDNA3.1 plasmid encoding
145 porcine GM-CSF (Dufour et al., 2000, Somasundaram et al., 1999) was kindly provided by
146 François Lefèvre (INRA, Jouy-en-Josas, France). The GM-CSF cassette was extracted by
147 *Apa* I and *Not* I digestion and inserted into the dephosphorylated pSINCP plasmid digested
148 with the same enzymes (Fig. 2A). pSINCP encoding porcine GM-CSF was characterized by
149 endonuclease restrictions and sequencing (not shown). Porcine PK15 cells were transfected
150 with 2 µg of pcDNA3.1/GM-CSF, pSINCP/GM-CSF, empty pcDNA3.1 or empty pSINCP as
151 described above. Forty-eight hours later, supernatants of each cell culture were collected and
152 evaluated for GM-CSF activity (Fig. 2B). This was done in vitro on TF-1 cells as previously

153 described (Loizel et al., 2005). In fact, these cells only grow in the presence of GM-CSF.
154 Briefly, TF-1 cells were incubated for 16 hours with different concentrations of recombinant
155 porcine GM-CSF (0-10 ng/ml) (R & D Systems, Minneapolis, MN, USA) or with the
156 supernatants collected above (final volume: 100 μ l). Each culture condition was prepared in
157 triplicate. Twenty μ l of Alamar Blue dye (Biosource International, Camarillo, CA, USA) were
158 then added. After 6 hours incubation, the optical densities (OD) of the cultures were read at
159 600 nm (original oxidized form of the dye) and at 570 nm (reduced form). The specific
160 absorbance of each culture corresponded to the difference between 600 nm OD and 570 nm
161 OD. Proliferation of the TF-1 cells was represented by the difference between the specific
162 absorbance of cells incubated with rGM-CSF or PK15 supernatants and that of non-stimulated
163 cells (= Δ specific absorbance).

164 These plasmids, or empty pcDNA3 or pSINCP plasmids were introduced into Escherichia
165 coli XL-1 blue strain, amplified and purified using the EndoFree plasmid Mega kit (Qiagen,
166 Hilden, Germany) according to the manufacturer's instructions.

167

168 *2.2 Animal experiments*

169 Two experiments were performed in pigs. In the first, 7 groups of 3 unvaccinated large white
170 pigs obtained from an air-filtered farm were housed and treated in accordance with the
171 regulations of the local veterinary office (Direction des Services Vétérinaires des Côtes
172 d'Armor, France). The pigs were intramuscularly injected in the neck 3 times at 2-week
173 intervals with 2 ml of plasmids DNA using 0.8 mm x 40 mm needles. The first injection was
174 administered when the pigs were 7 weeks old.

175 At each injection time, 600 μ g or 24 μ g of pcDNA3.1/P1-2A3C3D were co-injected
176 with 200 μ g of pcDNA3.1/GM-CSF in group 1 or group 2, respectively. Since pSINCP/P1-
177 2A3C3D is 1.7 times longer than pcDNA3.1/P1-2A3C3D, groups 3 and 4 were co-injected

178 with 1020 µg or 40 µg of pSINCP/P1-2A3C3D and 340 µg pSINCP/GM-CSF at each
179 injection time, respectively. Group 5 was co-injected with 600 µg of empty pcDNA3.1 and
180 200 µg of pcDNA3.1/GM-CSF. Group 6 was co-injected with 1020 µg empty-pSINCP and
181 340 µg pSINCP/GM-CSF. Group 7 was not injected throughout the assay.

182 In the second assay, 8 groups of 4 specific pathogen-free pigs were injected by IM
183 and/or ID routes 3 times at 3-week intervals. The first injection was administered when the
184 pigs were 7 weeks old. At each injection time, pigs were injected either with a total 600 µg of
185 pcDNA3.1/P1-2A3C3D + 200 µg of pcDNA3.1/GM-CSF or 1020 µg of pSINCP/P1-
186 2A3C3D + 340 µg pSINCP/GM-CSF. The ID injection was done in the dorsal surface of both
187 ears using 0.45 mm x 12 mm needles. It was controlled by (i) the parallel position of the
188 needle and the ear surface, (ii) the high pressure applied on the syringe to inject the solutions
189 and (iii) the transient generation of white spots. As in the first assay, groups 1 and 2 received
190 an IM injection (1 x 2 ml) of the pcDNA3.1 based or pSINCP based DNA vaccine,
191 respectively. Groups 3 and 4 received either an IM injection (half the DNA quantity, 1 ml in
192 each side of the animal) and an ID injection (half the DNA quantity, 0.25 ml on the top of
193 each ear) of the pcDNA3.1 based or pSINCP based DNA vaccine, respectively. Groups 5 and
194 6 were ID injected with the pcDNA3.1 based or pSINCP based DNA vaccine, respectively
195 (0.25 ml on the top of each ear). Two other groups were injected either with 600 µg of
196 pcDNA3.1/P1-2A3C3D + 200 µg of pcDNA3.1/GM-CSF or 1020 µg of pSINCP/P1-
197 2A3C3D + 340 µg of pSINCP/GM-CSF by both routes (IM + ID), respectively.

198 The pigs were observed for any adverse reaction after injection. Body temperature was
199 measured 4 hours after injection then daily. Relative daily weight gains were determined
200 (Stellmann et al., 1989) for each pig. Finally, the pigs were sacrificed at the end of the assay
201 and the injected region and other organs were examined to see whether the injection of
202 plasmids produced lesions on these organs.

203

204 *2.3 Determination of FMDV-specific serum antibodies*

205 Anti FMDV antibodies were first measured in pig sera using a commercial test: Ceditest
206 FMDV type O (Cedi Diagnostics B.V., Lelystad, The Netherlands) which is a blocking Elisa
207 (Chenard et al., 2003). Positive serums are those presenting 50% or more inhibition compared
208 to a high positive reference serum. In order to detect very low levels of antibodies, an Elisa
209 test was set up as follows: 96-well, flat-bottomed plates (Maxisorp; Nunc, Roskilde,
210 Denmark) were coated with a rabbit anti O1 BFS antiserum diluted in carbonate/bicarbonate
211 buffer pH 9.6 (Sigma, Saint Louis, MO, USA). Plates were blocked with PBS-tween20 0.05%
212 buffer supplemented with 10% bovine serum and 5% rabbit serum. The plates were then
213 incubated with FMDV antigen (O1 BFS) for 1 h at 37°C. After washings, samples of pig sera
214 diluted 1/40 in blocking buffer were added in duplicate to an antigen-coated well and to a
215 control well. After washings and incubation with a HRP (Horse Radish Peroxidase)
216 conjugated anti swine serum, the reaction was revealed with an OPD solution (Sigma).
217 Results were expressed as the difference in measured OD (optical density) between the
218 antigen coated and the control wells. A serum is considered as positive if the difference of OD
219 is superior to 0.20.

220

221 *2.4 FMDV neutralizing antibodies*

222 Neutralizing antibody assays were carried out in 96 wells as described in the OIE Manual of
223 Standards (OIE, 2000). Serial dilutions of sera were performed in duplicate and 50µl of each
224 were added for 1 h to 50µl of 100 TCID₅₀ of FMDV O1 BFS strain. Cell suspension was
225 then added to each well and the plates were incubated at 37°C for 3 days. The cells were fixed
226 with formalin and stained with methylene blue. Titres were expressed as the last serum
227 dilution that inhibited viral replication in 50% of the wells.

228

229 *2.5 Statistical analyses*

230 The data were analysed using the nonparametric Mann-Whitney test (Mann and Whitney,
231 1947) included in the Systat 9 software (Systat Software, Inc., Point Richmond, CA, USA).

232 This test was used as the generated data were few in number, did not present a normal
233 distribution and consisted of unpaired quantitative data.

234 The limit of significance was 0.05 for all comparisons.

235

236 **3. Results**

237 In vitro assays showed that each FMDV P1-2A3C3D construct and each GM-CSF construct
238 were effectively expressed in the porcine cell line PK15 (see Fig. 1 and 2 and Materials and
239 Methods). PK15 cells transfected with pSINCP/GM-CSF produced more GM-CSF than cells
240 transfected with pcDNA3.1/GM-CSF (Fig. 2). The immune potentials of the FMDV
241 constructs were assessed in swine. Serum samples from all pigs were analysed by ELISA and
242 viral neutralization assays for FMDV-specific and FMDV-neutralizing antibodies responses.

243 First, the capacity of a plasmid derived from the Sindbis virus as backbone to enhance
244 the induction of humoral immune response or to significantly reduce the quantity of plasmids
245 needed for the FMDV immunization was evaluated. An assay was performed to compare 2
246 different quantities of each of the 2 plasmid backbones (pcDNA3.1 and pSINCP). In the case
247 of pcDNA3.1, 600 µg of plasmids were injected in one group as in previous studies (Cedillo-
248 Barron et al., 2001) and 25 times fewer plasmids, i.e. 24 µg, were injected in another group in
249 accordance with the results with PrV DNA vaccination (Dory et al., 2005). Since the pSINCP
250 based construct is 1.7 times longer than the pcDNA3.1 based one, 1020 or 40 µg of
251 pSINCP/P1-2A3C3D were injected in two other groups in order to use the same number of
252 copies of molecules. The plasmids were injected by IM route since this was the one used for
253 the PrV-pSINCP study (Dory et al., 2005) and in many FMDV DNA vaccination studies (Guo
254 et al., 2005, Guo et al., 2004, Wong et al., 2002, Zhang et al., 2003). No fever, adverse
255 reaction or modification of the daily weight gains was observed in any group after any
256 injection. The results show that these vaccines were well tolerated by the animals. The
257 production of specific antibodies against FMDV was assessed by ELISA. No antibodies were
258 detected in the groups injected with the smaller quantities of FMDV P1-2A3C3D encoding
259 plasmids or in the control groups (not shown). No antibodies were detected in pigs injected
260 with 600 µg of pcDNA3.1/P1-2A3C3D + pcDNA3.1/GM-CSF (Table 1). Seven days after the

261 second injection, low levels of antibodies were detected in the group injected with 1020 µg of
262 pSINCP/P1-2A3C3D + pSINCP/GMCSF and antibody production was detected in all 3 pigs
263 14 days after the last injection. Extremely low and non-significant levels of neutralizing
264 antibodies were detected in the groups injected with high quantities of FMDV P1-2A3C3D
265 encoding plasmids. Interestingly, neutralizing antibodies were detected transiently or until the
266 end of the assay in 1 out of 3 pigs in the groups injected with 24 µg of pcDNA3.1/P1-
267 2A3C3D + pcDNA3.1/GM-CSF or 40 µg of pSINCP/P1-2A3C3D + pSINCP/GM-CSF,
268 respectively. Nevertheless, collectively these data show that the pSINCP and the pcDNA3.1
269 based DNA vaccine injected via the IM route induced a low production of FMDV specific
270 and neutralizing antibodies. Except for the production of specific antibodies, where the
271 pSINCP-based DNA vaccine induced stronger responses, there were no differences between
272 the 2 kinds of plasmids. No lesions were apparent in any of the organs observed during
273 necropsy of the animals at the end of the assay.

274 In a second assay, the influence of the route of administration of the DNA vaccines was
275 studied. In this context, and based on several papers published with this FMDV P1-2A3C3D
276 construct (Cedillo-Barron et al., 2001, Li et al., 2006, Niborski et al., 2006), ID and IM
277 injections were compared. As no specific antibodies were detected in the group injected with
278 40 µg of pSINCP/P1-2A3C3D, whereas all pigs in the group injected with 1020 µg of
279 plasmids were positive, the second study was limited to the injection of the largest quantity of
280 plasmids used above. Plasmids encoding GM-CSF were co-injected in all pigs. As in the first
281 assay, no fever, adverse reaction or modification of the daily weight gains was observed after
282 any injection in any of the animal groups. No FMDV specific and neutralizing antibodies
283 were detected in the groups injected with empty pcDNA3, empty pSINCP or in non-injected
284 pigs (data not shown). As in the first assay, no or few antibodies were detected in the group
285 injected by IM route (Table 2 and Fig. 3). Three out of 4 pigs developed antibodies against

286 FMDV 14 days after the third ID injection of pcDNA3.1/P1-2A3C3D plasmid (Table 2). The
287 4th pig was found positive 1 week later. All pigs injected with the pcDNA3.1 based DNA
288 vaccine by ID + IM routes were found positive two weeks after the third injection. For the
289 groups injected with pSINCP/P1-2A3C3D, only 2 out of 4 pigs were detected positive in the
290 IM + ID injected group. No positive pigs were detected in the IM or ID injected groups.
291 As in the first assay, neutralizing antibodies against FMDV were first detected after the
292 second injection (Fig. 3). IM injections resulted in the production of low neutralizing
293 antibodies titres with both plasmid backbones (Fig. 3A and 3B). ID injection resulted in a
294 significant increase of the production of neutralizing antibodies when pcDNA3.1/P1-2A3C3D
295 was used (Fig. 3A) but not when pSINCP/P1-2A3C3D was used (Fig. 3B). The pcDNA3.1
296 based DNA vaccine induced a significantly higher production of neutralizing antibodies than
297 the pSINCP based vaccine (Fig. 3C). On the other hand, IM + ID injection did not result in a
298 significant increase of the production of neutralizing antibodies in either case, compared to
299 the group injected by IM route only. No lesions were apparent in any of the organs observed
300 during necropsy of the animals at the end of the assay.

301

302

303 **4. Discussion**

304 Vaccination is one of the most important strategies used to control FMDV infection
305 (Grubman and Baxt, 2004, Sobrino et al., 2001). Although vaccination with inactivated
306 viruses has been shown to be efficient, it is associated with several problems related to safety
307 and to discrimination between vaccinated and infected animals. These disadvantages might be
308 overcome by using DNA vaccination. Several strategies designed to increase the efficacy of
309 DNA vaccines have been evaluated. For example, GM-CSF (Cedillo-Barron et al., 2001), IL-
310 18 (Mingxiao et al., 2007), C3d (Fan et al., 2007), IL-2 (Wong et al., 2002) or electroporation
311 (Kim et al., 2006) were described as promising adjuvants or strategies to increase vaccination
312 efficacy whereas several other strategies failed (Bergamin et al., 2007, Guo et al., 2004).
313 Despite this progress, and as is the case for the FMDV P1-2A3C3D construct used here, 3
314 injections of plasmids are often needed (Cedillo-Barron et al., 2001, Li et al., 2006, Xiao et
315 al., 2007). Under these conditions, DNA vaccination is not expected to be useful to protect
316 pigs in the case of a FMDV outbreak. Other strategies, that generated promising results in
317 other models, should therefore be tested for FMDV DNA vaccination. This is the case for a
318 new generation of non-replicative plasmids derived from the Sindbis virus that amplifies
319 transcription of the replicons encoded by this plasmid (Dubensky et al., 1996). Use of this
320 plasmid was able, for example, to enhance protection against a lethal Herpes Simplex Virus
321 infection (Hariharan et al., 1998) or to decrease the quantity of plasmids needed for
322 immunization against *Mycobacterium tuberculosis* (Kirman et al., 2003) or a lethal PRV
323 (Dory et al., 2005) infection. The potential beneficial effect of this kind of strategy in DNA
324 vaccination against FMDV has not been examined before, and was the aim of the present
325 study. The FMDV P1-2A3C3D cassette was inserted into the pSINCP plasmid and the ability
326 of this construct to induce the production of FMDV antibodies was compared with that of the
327 pcDNA3.1-based DNA vaccine. As in the original study (Cedillo-Barron et al., 2001, Li et al.,

2006), a plasmid encoding porcine GM-CSF was co-injected. As in previous studies using pSINCP-based DNA vaccines (Hariharan et al., 1998, Leitner et al., 2000a), the DNA vaccines were injected by IM route in an initial study. IM injection of the pcDNA3.1 based DNA vaccine led to no or a low production of FMDV-specific or neutralizing antibodies respectively. The production of neutralizing antibodies in particular was below that obtained by Cedillo-Barron with similar quantities of the same pcDNA3.1-based construct (Cedillo-Barron et al., 2001), when the plasmids were injected by IM + ID routes. IM injection of the pSINCP based DNA vaccine induced low or non-significant amounts of neutralizing antibodies. For each DNA vaccine, a 25-fold decrease of the quantity of plasmids resulted in a very low induction of FMDV antibodies. For some samples, neutralizing antibodies were found at a titer lower than 6, whereas no specific antibodies were detected by Elisa. An explanation of these discrepancies could be that the level of neutralising antibodies is too low, and maybe non significant. In fact, a special attention has to be made when interpreting low neutralisation titers (Chenard et al., 2003). Furthermore, for some other samples, specific but no neutralising antibodies were detected. All the discrepancies observed could be due to the fact that the Elisa and the virus neutralisation test are detecting different populations of antibodies (Chenard et al., 2003). It can be hypothesized from these results that the IM route of injection is not the best one to induce FMDV humoral immune responses. Thus a second assay using IM and/or ID routes of injection was performed. ID or ID+IM injections of the pSINCP-based DNA vaccine again induced a low production of FMDV antibodies. In contrast, ID injection of the pcDNA3.1-based DNA vaccine induced the production of FMDV-specific antibodies in 4 out of 4 pigs and significantly higher titres of neutralizing antibodies than the IM injection, as early as 7 days after the second injection of plasmids. The ID route was shown here to be the main one to induce production of FMDV-specific neutralizing antibodies. DNA vaccines applied to the surface of the skin have been described

353 to mainly induce the production of a Th2 immune response in different disease models (Hahn
354 et al., 2004, Zhu et al., 2004). This may be due to the presence of Langerhans or other
355 dendritic cells in the dermis (Leitner et al., 2000b, Peachman et al., 2003, Raz et al., 1994).
356 Furthermore, the fact that the concentration of the injected plasmids was 4 times higher in the
357 ID group (1200 µg/ml) than in the IM one (300 µg/ml) may also explain the observed
358 discrepancies. Finally, due to the properties of the pSINCP plasmid (Dubensky et al., 1996),
359 GM-CSF can potentially be produced at a high toxic concentration (Serafini et al., 2004) in
360 animals injected with the pSINCP/GM-CSF construct. The results presented here also suggest
361 that the dermal dendritic cells are more efficiently transfected with pcDNA3.1-based
362 plasmids than with pSINCP-based plasmids. This might be due to the larger size of the
363 pSINCP-based DNA vaccine (16306 bp vs 9829 bp for the pcDNA3.1-based construct) (Yin
364 et al., 2005). Some neutralizing antibodies were detected after the IM injection. It is
365 hypothesized that transfected myocytes (Leitner et al., 2000b) may also indirectly induce the
366 production of antibodies or that non-muscular cells might have been transfected after blood
367 transport of some plasmid molecules (Gravier et al., 2007).

368 At this stage, no conclusions can be drawn about the protective ability of the pSINCP/P1-
369 2A3C3D based DNA vaccine. In fact, there are descriptions in the literature of protection in
370 the presence of low titres (Sobrino et al., 2001) or even in the absence (Borrego et al., 2006)
371 of FMDV neutralizing antibodies. The protective efficacy of this DNA vaccine can only be
372 evaluated experimentally. Nevertheless, with the same FMDV antigens used, the level of
373 neutralizing antibodies was significantly much lower with the pSINCP-based DNA vaccine
374 than with the pcDNA3.1-based one previously shown to induce partial protection against
375 FMDV infection (Cedillo-Barron et al., 2001). Therefore, the possibility that the pSINCP-
376 based vaccine confers partial (or no) protection is strong. It was therefore decided not to
377 perform a FMDV challenge, which is the sole way of measuring the strength of a FMDV

378 vaccine, in order to prevent the pigs from useless suffering. Furthermore, ID injection of the
379 pcDNA3.1 based DNA vaccine was shown here to be the best way of inducing the production
380 of FMDV-specific neutralizing antibodies. It would be very interesting to evaluate the
381 protective potential of this DNA-vaccine injected by ID route after the second, or even first,
382 injection of plasmids.

383 In conclusion and contrary to our initial hypothesis, the pSINCP plasmid was shown to
384 be unable to enhance the production of FMDV neutralizing antibodies of the DNA vaccine
385 against FMDV encoding P1-2A3C3D. Furthermore, the route of injection of the plasmids was
386 shown to be essential for the induction of immune responses as the ID route showed higher
387 production of neutralizing antibodies than the intramuscular one.

388

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400 **References**

- 401
402 Bergamin, F., Saurer, L., Neuhaus, V., McCullough, K.C., Summerfield, A., 2007. Porcine B-
403 cell activating factor promotes anti-FMDV antibodies in vitro but not in vivo after
404 DNA vaccination of pigs. *Vet Immunol Immunopathol* 120, 115-123.
- 405 Borrego, B., Fernandez-Pacheco, P., Ganges, L., Domenech, N., Fernandez-Borges, N.,
406 Sobrino, F., Rodriguez, F., 2006. DNA vaccines expressing B and T cell epitopes can
407 protect mice from FMDV infection in the absence of specific humoral responses.
408 *Vaccine* 24, 3889-3899.
- 409 Cedillo-Barron, L., Foster-Cuevas, M., Belsham, G.J., Lefevre, F., Parkhouse, R.M., 2001.
410 Induction of a protective response in swine vaccinated with DNA encoding foot-and-
411 mouth disease virus empty capsid proteins and the 3D RNA polymerase. *J Gen Virol*
412 82, 1713-1724.
- 413 Cedillo-Barron, L., Foster-Cuevas, M., Cook, A., Gutierrez-Castaneda, B., Kollnberger, S.,
414 Lefevre, F., Parkhouse, R.M., 2003. Immunogenicity of plasmids encoding T and B
415 cell epitopes of foot-and-mouth disease virus (FMDV) in swine. *Vaccine* 21, 4261-
416 4269.
- 417 Chen, L., Shao, H., 2006. Extract from *Agaricus blazei* Murill can enhance immune responses
418 elicited by DNA vaccine against foot-and-mouth disease. *Vet Immunol Immunopathol*
419 109, 177-182.
- 420 Chenard, G., Miedema, K., Moonen, P., Schrijver, R.S., Dekker, A., 2003. A solid-phase
421 blocking ELISA for detection of type O foot-and-mouth disease virus antibodies
422 suitable for mass serology. *J Virol Methods* 107, 89-98.
- 423 Cox, S.J., Aggarwal, N., Statham, R.J., Barnett, P.V., 2003. Longevity of antibody and
424 cytokine responses following vaccination with high potency emergency FMD
425 vaccines. *Vaccine* 21, 1336-1347.
- 426 Dory, D., Torché, A.M., Béven, V., Blanchard, P., Loizel, C., Cariolet, R., Jestin, A., 2005.
427 Effective protection of pigs against lethal Pseudorabies virus infection after a single
428 injection of low-dose Sindbis-derived plasmids encoding PrV gB, gC and gD
429 glycoproteins. *Vaccine* 23, 3483-3491.
- 430 Du, Y., Jiang, P., Li, Y., He, H., Jiang, W., Wang, X., Hong, W., 2007. Immune responses of
431 two recombinant adenoviruses expressing VP1 antigens of FMDV fused with porcine
432 granulocyte macrophage colony-stimulating factor. *Vaccine* 25, 8209-8219.
- 433 Dubensky, T.W., Jr., Driver, D.A., Polo, J.M., Belli, B.A., Latham, E.M., Ibanez, C.E.,
434 Chada, S., Brumm, D., Banks, T.A., Mento, S.J., Jolly, D.J., Chang, S.M., 1996.
435 Sindbis virus DNA-based expression vectors: utility for in vitro and in vivo gene
436 transfer. *J Virol* 70, 508-519.
- 437 Dufour, V., Chevallier, S., Cariolet, R., Somasundaram, S., Lefèvre, F., Jestin, A., Albina, E.,
438 2000. Induction of porcine cytokine mRNA expression after DNA immunization and
439 pseudorabies virus infection. *J Interferon Cytokine Res* 20, 885-890.
- 440 Fan, H., Tong, T., Chen, H., Guo, A., 2007. Immunization of DNA vaccine encoding C3d-
441 VP1 fusion enhanced protective immune response against foot-and-mouth disease
442 virus. *Virus Genes* 35, 347-357.
- 443 Gravier, R., Dory, D., Laurentie, M., Bougeard, S., Cariolet, R., Jestin, A., 2007. In vivo
444 tissue distribution and kinetics of a pseudorabies virus plasmid DNA vaccine after
445 intramuscular injection in swine. *Vaccine* 25, 6930-6938.
- 446 Grubman, M.J., Baxt, B., 2004. Foot-and-mouth disease. *Clin Microbiol Rev* 17, 465-493.

- 447 Guo, H., Liu, Z., Sun, S., Bao, H., Chen, Y., Liu, X., Xie, Q., 2005. Immune response in
448 guinea pigs vaccinated with DNA vaccine of foot-and-mouth disease virus O/China99.
449 *Vaccine* 23, 3236-3242.
- 450 Guo, H.C., Liu, Z.X., Sun, S.Q., Leng, Q.W., Li, D., Liu, X.T., Xie, Q.G., 2004. The effect of
451 bovine IFN-alpha on the immune response in guinea pigs vaccinated with DNA
452 vaccine of foot-and-mouth disease virus. *Acta Biochim Biophys Sin (Shanghai)* 36,
453 701-706.
- 454 Hahn, U.K., Alex, M., Czerny, C.P., Bohm, R., Beyer, W., 2004. Protection of mice against
455 challenge with *Bacillus anthracis* STI spores after DNA vaccination. *Int J Med*
456 *Microbiol* 294, 35-44.
- 457 Hariharan, M.J., Driver, D.A., Townsend, K., Brumm, D., Polo, J.M., Belli, B.A., Catton,
458 D.J., Hsu, D., Mittelstaedt, D., McCormack, J.E., Karavodin, L., Dubensky, T.W., Jr.,
459 Chang, S.M., Banks, T.A., 1998. DNA immunization against herpes simplex virus:
460 enhanced efficacy using a Sindbis virus-based vector. *J Virol* 72, 950-958.
- 461 Kim, S.A., Liang, C.M., Cheng, I.C., Cheng, Y.C., Chiao, M.T., Tseng, C.J., Lee, F., Jong,
462 M.H., Tao, M.H., Yang, N S., Liang, S.M., 2006. DNA vaccination against foot-and-
463 mouth disease via electroporation: study of molecular approaches for enhancing VP1
464 antigenicity. *J Gene Med* 8, 1182-1191.
- 465 King, A.M., Underwood, B.O., McCahon, D., Newman, J.W., Brown, F., 1981. Biochemical
466 identification of viruses causing the 1981 outbreaks of foot and mouth disease in the
467 UK. *Nature* 293, 479-480.
- 468 Kirman, J.R., Turon, T., Su, H., Li, A., Kraus, C., Polo, J.M., Belisle, J., Morris, S., Seder, R.
469 A., 2003. Enhanced immunogenicity to *Mycobacterium tuberculosis* by vaccination
470 with an alphavirus plasmid replicon expressing antigen 85A. *Infect Immun* 71, 575-
471 579.
- 472 Leitner, W.W., Ying, H., Driver, D.A., Dubensky, T.W., Restifo, N.P. (2000a). Enhancement
473 of tumor-specific immune response with plasmid DNA replicon vectors. *Cancer Res*
474 60, 51-55.
- 475 Leitner, W.W., Ying, H., Restifo, N.P., 2000b. DNA and RNA-based vaccines: principles,
476 progress and prospects. *Vaccine* 18, 765-777.
- 477 Li, Y., Aggarwal, N., Takamatsu, H.H., Sterling, C.M., Voyce, C., Barnett, P.V., 2006.
478 Enhancing immune responses against a plasmid DNA vaccine encoding a FMDV
479 empty capsid from serotype O. *Vaccine* 24, 4602-4606.
- 480 Loizel, C., Blanchard, P., Grasland, B., Dory, D., Oger, A., Nignol, A.C., Cariolet, R., Jestin,
481 A., 2005. Effect of granulocyte-macrophage colony-stimulating factor on post-
482 weaning multisystemic wasting syndrome in porcine circovirus type-2-transfected
483 piglets. *Int J Exp Pathol* 86, 33-43.
- 484 Mann, H.B., Whitney, D.R., 1947. On a test of whether one of two random variables is
485 stochastically larger than the other. *Ann. Math. Statist.* 18, 50-60.
- 486 Mingxiao, M., Ningyi, J., Juan, L.H., Min, Z., Guoshun, S., Guangze, Z., Huijun, L., Xiaowei,
487 H., Minglan, J., Xu, L., Haili, M., Yue, J., Gefen, Y., Kuoshi, J., 2007.
488 Immunogenicity of plasmids encoding P12A and 3C of FMDV and swine IL-18.
489 *Antiviral Res* 76, 59-67.
- 490 Niborski, V., Li, Y., Brennan, F., Lane, M., Torche, A.M., Remond, M., Bonneau, M.,
491 Riffault, S., Stirling, C., Hutchings, G., Takamatsu, H., Barnett, P., Charley, B.,
492 Schwartz-Cornil, I., 2006. Efficacy of particle-based DNA delivery for vaccination of
493 sheep against FMDV. *Vaccine* 24, 7204-7213.
- 494 OIE, 2000. OIE Manual of Standards for diagnostics tests and vaccines, 4th edition edn.

495 Park, J.H., Kim, S.J., Oem, J.K., Lee, K.N., Kim, Y.J., Kye, S.J., Park, J.Y., Joo, Y.S., 2006.
496 Enhanced immune response with foot and mouth disease virus VP1 and interleukin-1
497 fusion genes. *J Vet Sci* 7, 257-262.

498 Peachman, K.K., Rao, M., Alving, C.R., 2003. Immunization with DNA through the skin.
499 *Methods* 31, 232-242.

500 Raz, E., Carson, D.A., Parker, S.E., Parr, T.B., Abai, A.M., Aichinger, G., Gromkowski, S.H.,
501 Singh, M., Lew, D., Yankauckas, M.A., Baird, S.M., Rhodes, G.H., 1994. Intradermal
502 gene immunization: The possible role of DNA uptake in the induction of cellular
503 immunity to viruses. *Proc. Natl. Acad. Sci. USA* 91, 9519-9523.

504 Rodrigo, M. J., Dopazo, J., 1995. Evolutionary analysis of the picornavirus family. *J Mol*
505 *Evol* 40, 362-371.

506 Ryan, M.D., Belsham, G.J., King, A.M., 1989. Specificity of enzyme-substrate interactions in
507 foot-and-mouth disease virus polyprotein processing. *Virology* 173, 35-45.

508 Serafini, P., Carbley, R., Noonan, K.A., Tan, G., Bronte, V., Borrello, I., 2004. High-dose
509 granulocyte-macrophage colony-stimulating factor-producing vaccines impair the
510 immune response through the recruitment of myeloid suppressor cells. *Cancer Res* 64,
511 6337-6343.

512 Sobrino, F., Saiz, M., Jimenez-Clavero, M.A., Nunez, J.I., Rosas, M.F., Baranowski, E., Ley,
513 V., 2001. Foot-and-mouth disease virus: a long known virus, but a current threat. *Vet*
514 *Res* 32, 1-30.

515 Somasundaram, C., Takamatsu, H., Andréoni, C., Audonnet, J.C., Fisher, L., Lefèvre, F.,
516 Charley, B., 1999. Enhanced protective response and immuno-adjuvant effects of
517 porcine GM-CSF on DNA vaccination of pigs against Aujeszky's disease virus. *Vet.*
518 *Immunol. Immunopathol.* 70, 277-287.

519 Stellmann, C., Vannier, P., Chappuis, G., Brun, A., Dauvergne, M., Frageaud, D., Bugand,
520 M., Colson, X., 1989. The potency testing of pseudorabies vaccines in pigs. A
521 proposal for a quantitative criterion and a minimum requirement. *J Biol Stand* 17, 17-
522 27.

523 *The Veterinary Record*, 2007. FMD outbreak linked to defective drain. *Vet Rec.* 161, 362-
524 363.

525 Wong, H.T., Cheng, S.C., Sin, F.W., Chan, E.W., Sheng, Z.T., Xie, Y., 2002. A DNA vaccine
526 against foot-and-mouth disease elicits an immune response in swine which is
527 enhanced by co-administration with interleukin-2. *Vaccine* 20, 2641-2647.

528 Xiao, C., Jin, H., Hu, Y., Kang, Y., Wang, J., Du, X., Yang, Y., She, R., Wang, B., 2007.
529 Enhanced protective efficacy and reduced viral load of foot-and-mouth disease DNA
530 vaccine with co-stimulatory molecules as the molecular adjuvants. *Antiviral Res* 76,
531 11-20.

532 Yin, W., Xiang, P., Li, Q., 2005. Investigations of the effect of DNA size in transient
533 transfection assay using dual luciferase system. *Anal Biochem* 346, 289-294.

534 Zhang, H.Y., Sun, S.H., Guo, Y.J., Zhou, F.J., Chen, Z.H., Lin, Y., Shi, K., 2003. Immune
535 response in mice inoculated with plasmid DNAs containing multiple-epitopes of foot-
536 and-mouth disease virus. *Vaccine* 21, 4704-4707.

537 Zhu, W., Thomas, C.E., Sparling, P.F., 2004. DNA immunization of mice with a plasmid
538 encoding *Neisseria gonorrhoea* PorB protein by intramuscular injection and epidermal
539 particle bombardment. *Vaccine* 22, 660-669.

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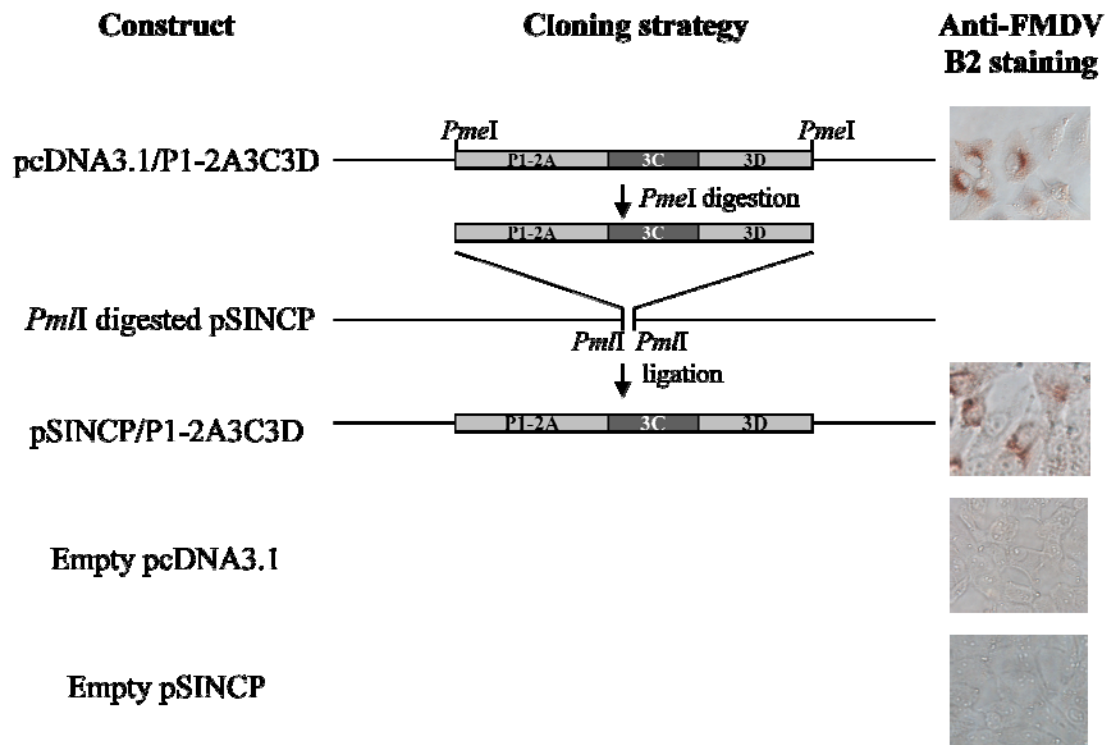


Fig. 1. Construction and characterization of the pSINCP/P1-2A3C3D construct.

pSINCP/P1-2A3C3D was constructed as described in Material and Methods. Porcine PK15 cells transfected with pcDNA3.1/P1-2A3C3D, pSINCP/P1-2A3C3D, empty pSINCP or non transfected cells were stained with anti-VP1 monoclonal antibody B2 24 hours after transfection.

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548 **Table 1 : Induction of anti-FMDV specific and neutralizing antibodies after intramuscular injection**
 549 **of different constructs.**

Group	Pig #		Day 0	Day 7	Day 14	Day 21	Day 28	Day 35	Day 42
				<i>Inj1+7d</i>	<i>Inj1+14d</i>	<i>Inj2 +7d</i>	<i>Inj2+14d</i>	<i>Inj3+7d</i>	<i>Inj3+14d</i>
pcDNA3.1/P1-2A3C3D 600 µg	1.1	O.D.	0.02	0.03	0.08	0.00	0.00	0.00	0.00
		NAb titer	0	0	4	0	0	0	0
	1.2	O.D.	0.02	0.00	0.00	0.05	0.15	0.00	0.00
		NAb titer	0	0	3	2	0	2	2
	1.3	O.D.	0.00	0.06	0.04	0.09	0.00	0.00	0.02
		NAb titer	0	0	0	3	2	3	2
pcDNA3.1/P1-2A3C3D 24 µg	1.4	O.D.	0.00	0.00	0.00	0.00	0.07	0.00	0.00
		NAb titer	0	0	2	3	2	0	0
	1.5	O.D.	0.06	0.07	0.08	0.04	0.04	0.11	0.00
		NAb titer	0	0	0	0	0	0	0
	1.6	O.D.	0.07	0.00	0.06	0.04	0.02	0.00	0.00
		NAb titer	0	0	0	0	0	0	0
pSINCP/P1-2A3C3D 1020 µg	1.7	O.D.	0.00	0.07	0.00	0.05	0.00	0.68	0.37
		NAb titer	0	0	0	0	3	0	3
	1.8	O.D.	0.06	0.12	0.00	0.48	0.28	1.02	1.00
		NAb titer	0	0	0	0	0	0	0
	1.9	O.D.	0.10	0.08	0.10	0.21	0.15	0.06	0.37
		NAb titer	0	0	0	0	4	2	6
pSINCP/P1-2A3C3D 40 µg	1.10	O.D.	0.03	0.04	0.00	0.00	0.06	0.04	0.04
		NAb titer	0	0	0	0	0	0	0
	1.11	O.D.	0.00	0.05	0.02	0.06	0.00	0.00	0.00
		NAb titer	0	0	0	0	0	0	0
	1.12	O.D.	0.10	0.08	0.07	0.10	0.10	0.02	0.00
		NAb titer	0	0	3	3	4	3	3
empty pcDNA3.1 600 µg	1.13	O.D.	0.01	0.00	0.00	0.00	0.04	0.03	0.00
		NAb titer	0	0	0	0	0	0	0
	1.14	O.D.	0.00	0.09	0.00	0.06	0.00	0.05	0.00
		NAb titer	0	0	0	0	0	0	0
	1.15	O.D.	0.14	0.04	0.10	0.09	0.08	0.00	0.00
		NAb titer	0	0	0	0	0	0	0
empty pSINCP 1020 µg	1.16	O.D.	0.02	0.00	0.00	0.00	0.00	0.00	0.00
		NAb titer	0	0	0	0	0	0	0
	1.17	O.D.	0.06	0.00	0.04	0.08	0.05	0.03	0.00
		NAb titer	0	0	0	0	0	0	0
	1.18	O.D.	0.00	0.00	0.06	0.02	0.00	0.00	0.00
		NAb titer	0	0	0	0	0	0	0

550
 551 Anti-FMDV specific and neutralizing antibodies titres were determined after injections of 600
 552 µg or 24 µg of pcDNA3.1/P1-2A3C3D, 1020 40 µg of pSINCP/P1-2A3C3D, 600 µg of
 553 empty pcDNA3.1 or 1020 µg of empty pSINCP. Specific antibodies titres were determined by
 554 ELISA as described in Materials and Methods. The optical densities are shown. Serums with
 555 optical densities superior to 0.20 are considered as positives. Anti-FMDV neutralizing
 556 antibodies titres were determined as described in Materials and Methods.

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 558 *Inj = injection; d = days*
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560 **Table 2: Induction of anti-FMDV specific antibodies after intramuscular and/or intradermal**
 561 **injection of different constructs.**

Group	Pig #	Day 0	Day 14	Day 21	Day 28	Day 35	Day 42	Day 49	Day 56	Day 63
			<i>Inj1+14d</i>	<i>Inj1+21d</i>	<i>Inj2+7d</i>	<i>Inj2+14d</i>	<i>Inj2+21d</i>	<i>Inj3+7d</i>	<i>Inj3+14d</i>	<i>Inj3+21d</i>
pcDNA3.1/P1-2A3C3D IM	2.1	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.2	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.3	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.4	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
pSINCP/P1-2A3C3D IM	2.5	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.6	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.7	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.8	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
pcDNA3.1/P1-2A3C3D IM+ID	2.9	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	52.5	<50.0
	2.10	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	57.4	54.4
	2.11	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	54.4	<50.0
	2.12	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	57.0	55.0
pSINCP/P1-2A3C3D IM+ID	2.13	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	63.4	52.3
	2.14	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	50.0	<50.0
	2.15	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.16	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
pcDNA3.1/P1-2A3C3D ID	2.17	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	52.9	<50.0
	2.18	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	50.0	68.2	59.6
	2.19	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	50.0	<50.0
	2.20	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	50.0
pSINCP/P1-2A3C3D ID	2.21	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.22	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.23	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.24	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
empty pcDNA3.1 IM+ID	2.25	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.26	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.27	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.28	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
empty pSINCP IM+ID	2.29	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.30	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.31	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0
	2.32	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0	<50.0

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Anti-FMDV specific antibodies productions were determined after IM, ID or IM+ID injections of pcDNA3.1/P1-2A3C3D or pSINCP/P1-2A3C3D, or after IM+ID injections of empty pcDNA3.1 or empty pSINCP. The level of these antibodies was measured using a blocking ELISA and are indicating as a % of inhibition of a positive control reference serum. Positive serums (Pos) are those presenting 50 % or more inhibition.

Inj = injection; d = days