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Monitoring of Venturia inaequalis harbouring the QoI resistance G143A mutation in French orchards as revealed by PCR assays

Séverine Fontaine, Florent Remuson, Laurence Fraissinet-Tachet, Annie Micoud, Roland Marmeisse and Delphine Melaya*

Abstract

BACKGROUND: Genetic resistance to QoI fungicides may account for recent failures to control Venturia inaequalis (Cooke) Winter in French orchards. Two PCR-based assays were developed to detect the G143A point mutation in the fungal mitochondrial cytochrome b gene. The mutation is known to confer a high level of resistance to QoI fungicides. Occurrence of the G143A mutation in French field isolates collected from 2004 to 2007 was monitored.

RESULTS: The QoI-resistant cytochrome b allele was specifically detected either following the cleavage of the amplified marker by a restriction endonuclease (CAPS assay) or its amplification using an allele-specific PCR primer. Using either method, the G143A mutation was found in 42% of the 291 field samples originating from French orchards in which apple scab proved difficult to be controlled. Monitoring of the G143A mutation in orchards located in 15 French administrative regions indicated that the mutation was detected at least once in nine of the regions, and its presence ranged from 33% to 64% of the orchards analysed in 2004 and in 2007 respectively.

CONCLUSION: The PCR-based methods developed in this study efficiently reveal the presence of the G143A mutation in French V. inaequalis field populations.

Keywords: Venturia inaequalis; apple scab; QoI; fungicide resistance; allele-specific PCR; molecular diagnostics

1 INTRODUCTION

Apple scab, caused by the fungus Venturia inaequalis (Cooke) Winter, is characterised by brown to olive-coloured spots on apple tree leaves and fruits. This major disease in apple tree orchards is mainly controlled through fungicide treatments. Quinone outside inhibiting (QoI) fungicides are one of the latest families of chemicals that play an important role in plant protection against many phytopathogenic fungi including V. inaequalis. These molecules specifically inhibit cell respiration by binding at the ubiquinol oxidation centre (Qo site) of the mitochondrial cytochrome bc1 complex.1,2

Resistance to QoI fungicides has been detected in several fungal pathogens such as Blumeria graminis Speer3 and Mycosphaerella fijiensis Morelet4 or Oomycetes such as Pseudoperonospora cubensis (Burk. & Curt.) Rostosev.5–7 QoI resistance in V. inaequalis was first reported in 1997 in European field and experimental trials.8 In France, the commercial QoI fungicides used to control V. inaequalis correspond to strobilurins. Although strobilurins have been successfully used on apple trees to control scab since 1998, the first cases of QoI resistance that occurred in French commercial apple tree orchards were reported in 2002 (unpublished results from the Service Régional Rhône-Alpes de la Protection des Végétaux, France). Several mechanisms leading to QoI resistance have been reported in V. inaequalis. One known mode of resistance is the metabolism of kresoxim-methyl, a commercial strobilurin molecule, by esterases and cytochrome P450 monooxygenases.9 Under laboratory conditions, induction of the alternative respiration pathway has also been observed during conidia germination.10 This latter phenomenon could possibly occur in apple tree orchards and contribute to resistance in the field.8 Finally, the commonly found target-site mutation referred to as the G143A mutation, which accounts for a high level of resistance to QoI in many plant pathogens, has also been identified in QoI-resistant V. inaequalis isolates. This point mutation occurs in the mitochondrial cytochrome b (Cyt b) gene and results in a change of glycine to alanine at position 143 of the protein.11 However, Steinfeld et al.12 obtained an isolate of V. inaequalis that
presented a high level of resistance to QoI without the G143A mutation from an experimental orchard in Switzerland. As the alternative oxidase pathway was not found to be overexpressed in this isolate, additional mutations in genes encoding components of the cytochrome b1 complex were suspected.

Monitoring of QoI resistance from field-collected infected material (leaves, apples) is of practical importance to evaluate the efficiency of these molecules and to reorient fungicide treatments in the corresponding orchards. Currently, detection of *V. inaequalis* QoI-resistant isolates relies on in vitro test systems based on spore germination on fungicide-amended media. As viability of *V. inaequalis* conidia recovered from commercial orchards treated with pesticides is usually low, alternative monitoring methods are needed. PCR-based detection is a valuable tool when the fungicide-resistant phenotype is clearly associated with a known point mutation. This is the case for the single nucleotide polymorphism that characterises the G143A mutation in the Cyt b gene of several fungal QoI-resistant pathogens including *V. inaequalis*.

The objectives of this study were to develop a rapid molecular test to detect the G143A mutation in field-collected apple scab lesions and to use this method to monitor this point mutation in apple tree orchards in France. Two methods based on the amplification of a portion of the Cyt b gene were developed. The first one requires the digestion of PCR products with a restriction enzyme that specifically cleaves the amplified fragment at the site of the mutation. The second one is an allele-specific PCR detection using an oligonucleotide primer that specifically binds to the mutant sequence.

## 2 MATERIALS AND METHODS

### 2.1 Fungal isolates

Two strains of *V. inaequalis* were isolated from infected apple tree leaves and maintained in pure culture on potato dextrose agar medium (Table 1). The QoI-sensitive single-spore isolate SFR2, collected in July 2002 originated from an orchard that had never been treated with fungicides. Single-spore isolate 160, collected in June 2003 from a commercial orchard in south France, displayed a high resistance factor (RF = 283) towards kresoxim-methyl (see Section 2.4). In addition to *V. inaequalis* isolates, strains of *Botrytis cinerea* Pers., *Rhizopus* sp. and *Penicillium* sp. were used to test for the specificity of the PCR primers (Table 1).

A total of 317 field samples were collected from 2004 to 2007, and 291 were analysed (see supporting information in Table S1). They originated from commercial and experimental French orchards where strobilurin treatments were suspected to be ineffective to control the pathogen. Infected apple leaves collected from orchards that had never been treated with any fungicides were used as control QoI-sensitive field samples.

For each orchard, *V. inaequalis* spores were collected from 30–50 lesion-bearing leaves or fruits with a brush. Spore suspensions were adjusted in distilled water to 1 × 10^9 conidia mL^-1. For each orchard, 30 lesions on leaves (ca 0.6 × 0.6 cm) or fruits (ca 100 mg) were collected (1 lesion per leaf or fruit), pooled and stored at −20 °C until DNA extraction. In addition, individual leaf lesions were frozen separately. The latter samples were used as starting material to evaluate the detection thresholds of both PCR methods.

### 2.2 DNA extraction

Frozen mycelia as well as mixed and unique lesion samples were ground in liquid nitrogen using a mortar and pestle. Fungal DNA or DNA from a plant–fungus mixture was extracted and purified using Nucleospin Plant DNA extraction kit (Macherey Nagel) according to the manufacturer’s recommendations. DNA was resuspended in 100 μL of elution buffer and quantified using a NanoDrop ND-1000 spectrophotometer.

| Table 1. Geographic origin and sampling year of the fungal isolates used in this study |
|---------------------------------|--------------------------------|
| **Isolates** | **Sampling year** | **French administrative region (locality)** | **EC50**(mg L⁻¹) |
| *Venturia inaequalis* | 2002 | Rhône-Alpes (Saint Marcel d’Urfe) | 0.06 |
| 160 | 2003 | Midi-Pyrénées (Montaubean) | 8.5 |
| *Botrytis cinerea* | 2003 | Rhône-Alpes (Saint Genis Laval) | nd³ |
| *Rhizopus* sp. | 2003 | Rhône-Alpes (Saint Didier sous Riverie) | nd³ |
| *Penicillium* sp. | 2003 | Rhône-Alpes (Saint Didier sous Riverie) | nd³ |

*a* EC50 refers to the concentration of kresoxim-methyl that leads to a 50% inhibition of the mycelia growth rate.

³ nd = not determined.

### 2.3 PCR-based detection

#### 2.3.1 Cleaved amplified polymorphic sequence (CAPS) assay

The PCR primers PS1 and PR1 (Table 2) were designed using the published sequence of the *V. inaequalis* Cyt b gene (GenBank accession number AF004559). They hybridise in exons 3 and 4 of the gene, respectively, and amplify a 488 bp fragment containing the region of interest, i.e. codon 143 site of the G143A mutation. This point mutation changes the sequence 5'−GGTGC−3' into 5'−GCTGC−3', which is specifically recognised by the restriction endonuclease *Fnu4HI*.

PCR amplifications were performed in a 25 μL volume containing 1 μL of DNA (1−10 ng), 0.5 μM of each primer, 200 μM of each dNTP, 0.5 mM magnesium chloride and 2 μL of Taq-DNA polymerase in the appropriate buffer (Invitrogen). An initial denaturation step at 94 °C for 3 min was followed by 35 cycles that each included a denaturation at 94 °C for 30 s, a primer annealing step at 55 °C for 45 s and a polymerisation step at 72 °C for 50 s. Amplifications were terminated by a 5 min final extension at 72 °C. PCR products from the sensitive (SFR2) and resistant (160) isolates were directly sequenced using the BigDye3.1 Terminator sequencing kit (Applied Biosystems) and analysed on an ABI3730XL capillary sequencer (Applied Biosystems) at Genoscreen Services (Lille, France). A quantity of 10 μL of the PCR products was digested overnight by 1.5 U of *Fnu4HI* (New England Biolabs) at 37 °C. Restriction fragments were separated on 2.5% agarose gels in 0.5 × TBE and stained with ethidium bromide. Gels were visualised using a UV transilluminator and photographed using the Gel Doc 2000 apparatus (BioRad).

#### 2.3.2 Allele-specific PCR (AS-PCR)

Allele-specific PCR primers that would preferentially bind to the mutant sequence were designed. The backbone sequence of these primers corresponds to primer G143AMM0 for which the 3’ G nucleotide is complementary to the C residue specific to the
Table 2. Primers used in this study. For the allele-specific primers G143AMM0 to G143AMM4, the introduced mismatch bases relative to the Venturia inaequalis Cyt b wild-type sequence are underlined

<table>
<thead>
<tr>
<th>Name</th>
<th>Orientation</th>
<th>Sequence (5′ → 3′)</th>
<th>Nucleotide positiona</th>
</tr>
</thead>
<tbody>
<tr>
<td>PS1</td>
<td>Forward</td>
<td>GTTACACGCTCCTCGGGTTAT</td>
<td>5025–5045</td>
</tr>
<tr>
<td>PR1</td>
<td>Reverse</td>
<td>AGGCCCTCCCAAGAATTAG</td>
<td>5513–5495</td>
</tr>
<tr>
<td>G143AMM0</td>
<td>Reverse</td>
<td>GGGTTTGATGACAGTTGTGAG</td>
<td>5457–5437</td>
</tr>
<tr>
<td>G143AMM1</td>
<td>Reverse</td>
<td>GGGTTTGATGACAGTGCTG</td>
<td>5457–5437</td>
</tr>
<tr>
<td>G143AMM2</td>
<td>Reverse</td>
<td>GGGTTTGATGACAGTAAGAG</td>
<td>5457–5437</td>
</tr>
<tr>
<td>G143AMM3</td>
<td>Reverse</td>
<td>GGGTTTGATGACAGTTGAAG</td>
<td>5457–5437</td>
</tr>
<tr>
<td>G143AMM4</td>
<td>Reverse</td>
<td>GGGTTTGATGACAGTGAG</td>
<td>5457–5437</td>
</tr>
<tr>
<td>PS-exon7</td>
<td>Forward</td>
<td>GGTGTTATACGTATGTTGACGTA</td>
<td>10394–10418</td>
</tr>
<tr>
<td>PR-exon7</td>
<td>Reverse</td>
<td>CAAGAGTCAATTCTACAAAGTGAG</td>
<td>10631–10605</td>
</tr>
</tbody>
</table>

a Position as referred to the GeneBank accession AF004559.

mutant allele (Table 2). Enhanced allelic discrimination was tested by introducing an additional mismatch in the 3′ subterminal region of the primer, thus creating primers G143AMM1, G143AMM2, G143AMM3 and G143AMM4 (Table 2). These allele-specific primers were used in combination with primer PS1 and tested to amplify a 433 bp DNA fragment.

All AS-PCR reactions included, as an amplification control, a second set of primers, namely PS-exon7 and PR-exon7 (Table 2). This primer pair allows, from any V. inaequalis Cyt b alleles, the amplification of a 238 bp sequence located some 5 kb downstream of the G143 mutation. PCR amplifications were performed in a 25 µL volume containing 0.2 µM of allele-specific primers and 0.1 µM of control primers (PS-exon7 and PR-exon7). Other AS-PCR components included 1 µL of DNA (1–10 ng), 400 µM of each dNTP, 0.5 mM of magnesium chloride and 2 U of Taq-DNA polymerase in the appropriate buffer (Invitrogen). An initial denaturation at 94 °C for 3 min was followed by 30 cycles that each included denaturation at 94 °C for 30 s, a primer annealing step at 60 °C for 45 s, and extension at 72 °C for 50 s. Amplifications were followed by a final extension of 3 min at 72 °C. PCR products were separated on 1% agarose gels in 0.5 × TBE and stained with ethidium bromide.

3.1 PCR-based detection of the G143A mutation in Venturia inaequalis

3.1.1 Cleaved Amplified polymorphic sequence (CAPS) assay

The preliminary set of this experimental work consisted of controlling the primer specificity towards V. inaequalis. Primer pair PS1–PR1 amplified a 488 bp fragment from DNA samples extracted from pure cultures of both QoI-sensitive (isolate SFR2) and resistant (isolate 160) V. inaequalis mycelia, as well as from DNA extracted from V. inaequalis-infected apple leaves and fruits collected in orchards. No amplification was obtained using DNA from non-infected apple tree leaves or other fungi such as Botrytis cinerea, Penicillium spp. and Rhizopus sp. (data not shown).

Sequences of the PCR products from the sensitive and resistant isolates were identical to the published sequence of V. inaequalis Cyt b gene, except for a guanine to cytosine transversion in the sequence from the isolates 160 responsible for the typical G143A mutation conferring QoI resistance (Fig. 1). This mutation created an Fnu4HI (5′-GCTGC-3′) restriction site in the mutant sequence. Upon digestion with this endonuclease, two DNA fragments of 412 and 76 bp were obtained with the PCR fragment from the resistant isolate 160, whereas, as expected, the PCR product from the sensitive isolate SFR2 was not cleaved (Fig. 2), allowing CAPS analysis on composite samples. As a control for population analysis, DNA amplified from lesions collected in several control orchards that had never been treated with strobilurins failed to control apple scab were frequently partially digested by Fnu4HI. On the other hand, DNA amplified from numerous samples collected in orchards in which strobilurin treatments were not effective contained Fnu4HI digestion sites (Fig. 2). Partial digestions probably resulted from the coamplification of sensitive and resistant Cyt b alleles, as the field-
PCR-based detection of QoI resistance in Venturia inaequalis

3.1.2 Allele-specific PCR (AS-PCR)

In order to improve the sensitivity of the PCR detection method, the authors developed an AS-PCR method. For this purpose, primer PS1, which should hybridise to any V. inaequalis Cyt b allele, was combined with a second primer designed to bind specifically to the sequence of the mutant allele conferring QoI resistance. The first AS primer designed, G143AMM0 (Table 2), differed from the wild-type sequence by having a G at its 3′ end, which corresponds to the G143A mutation. Using the primer pair PS1-G143AMM0, both wild-type and mutant alleles were equally amplified at either annealing temperature tested (data not shown). Therefore, four additional AS primers that differed from G143AMM0 by a second mismatch in position −1, −2 or −3 relative to the 3′ end of the primer were designed (Table 2). In combination with primer PS1, these four primers amplified a 433 bp fragment from both sensitive and resistant isolates at an annealing temperature of 55 °C. Allelic discrimination, i.e. specific amplification of the QoI-resistant allele, was achieved using any of the four primer pairs at the annealing temperature of 60 °C (Fig. 4). Primer G143AMM1 was then chosen as the AS primer for all subsequent experiments. To make sure that the absence of amplification of a 433 bp fragment did not result from PCR inhibition, all AS-PCR reactions included a second primer pair (PS-exon7 and PR-exon7), which allowed the amplification of a 238 bp fragment from all Cyt b alleles (Fig. 4).

The sensitivity of the AS-PCR protocol was evaluated by amplifying Cyt b mutant allele DNA molecules when mixed at different ratios with wild-type allele molecules. The mutant allele was amplified at the lowest ratio tested, 1% of DNA extracted from leaves infected with a QoI-resistant isolate to 99% of DNA extracted from leaves infected with a QoI-sensitive isolate (Fig. 5), thus showing a higher sensitivity of the AS-PCR detection method compared with CAPS analysis.

3.2 Spore germination assay versus molecular detection methods

Among the 317 samples collected from orchards from 2004 to 2007, only 284 samples were analysed for QoI resistance by both the bioassay and the PCR approaches. The 33 remaining samples were only tested by PCR approaches. Here, 74% of spore germination assays could not be interpreted owing to an excessively low quantity of spores collected from the leaves or

![Image](https://example.com/image1)

**Figure 1.** Partial alignment of the cytochrome b sequences from the QoI-sensitive (SFR2) and resistant (160) Venturia inaequalis isolates to the GeneBank AF004559 sequence. Note the G to C transversion at nucleotide position 5437 of the Cyt b gene, leading to the amino acid change glycine to alanine responsible for QoI resistance.

![Image](https://example.com/image2)

**Figure 2.** Fnu4Hl digestion of the 488 bp PCR fragment of the cytochrome b gene generated by primer pair PS1–PR1 using DNA extracted from QoI-resistant Venturia inaequalis isolate 160 (lane 2), QoI-sensitive isolate SFR2 (lane 3) and apple scab lesions collected in an orchard where resistant isolates predominated (r; lane 4) or were absent (s; lane 5). MW, 1 kb DNA ladder (Fermentas).

![Image](https://example.com/image3)

**Figure 3.** Sensitivity of the CAPS method. DNA extracted from individual lesions caused by QoI-resistant (R) and QoI-sensitive (S) isolates were mixed at different ratios (from 1:99 to 40:60, w: w) before amplification of a Cyt b 488 bp DNA fragment which was digested with Fnu4Hl. The 100% sensitive and 100% resistant corresponded to DNA extracted from mycelium of the QoI-sensitive isolate SFR2 and the QoI-resistant isolate 160, respectively. MW, 1 kb DNA ladder (Fermentas).

![Image](https://example.com/image4)

**Figure 4.** Allele-specific PCR amplification of the G143A Cyt b allele. DNA extracted from Venturia inaequalis QoI-resistant (a) and sensitive (b) isolates were used as templates for the PCR amplifications which used primer PS1 in combination with one of the five allele-specific primers G143AMM0 to G143AMM4 that specifically bind to the mutant cytochrome b sequence. As a positive amplification control, primers PS-exon7 and PR-exon7, which amplify a 238 bp fragment, were included in all PCR reactions. Primer G143AMM0, which differs from the wild-type sequence by one nucleotide at its 3′ end, was not specific at an annealing temperature of 60 °C. MW, 1 kb DNA ladder (Fermentas).
to a very low germination rate on fungicide-free medium, although an unusually low threshold value of 10% was used. In contrast, only in 8% of these samples did PCR amplification of the Cyt b fragment fail. This could be ascribed to degradation of the DNA extracted from excessively old lesion tissues or leaf samples not well preserved between their collection and delivery in the laboratory.

The QoI-sensitive isolate SFR2 showed 50% mycelia growth inhibition when using 0.06 mg L\(^{-1}\) of kresoxim-methyl (Table 1). Comparatively, QoI-resistant isolate 160 exhibited an EC\(50\) for mycelium of 8.5 mg L\(^{-1}\) of Qo fungicide. The present data are consistent with the baseline sensitivity (0.09 mg L\(^{-1}\)) and the mean EC\(50\) value (6.09 mg L\(^{-1}\)) found in V. inaequalis populations exposed to kresoxim-methyl in Quebec orchards.\(^{14}\)

Using kresoxim-methyl, the EC\(50\) for spores collected in French orchards that had never been treated with strobilurin varied between 0.001 and 0.006 mg L\(^{-1}\) (Table 3). These EC\(50\) values are in accordance with literature data found for sensitive isolates (0.0006–0.007 mg L\(^{-1}\)).\(^{6,12}\) As illustrated in Fig. 6, spores collected in 46 of the 73 analysed orchards had EC\(50\) values that were at least 10 times higher (up to more than 10\(^4\) times) than the values recorded for sensitive fungal isolates. For the resistant groups of orchards that were considered from in vitro germination tests as being infected by QoI-resistant isolates, molecular analyses made it possible to detect the presence of the G143A mutation in 35 (76%) of them. For the remaining 11 resistant orchards, the G143A mutation could not be detected by a PCR approach, in spite of the fact that their EC\(50\) and MIC values were respectively 10–10\(^0\)0 times higher than values recorded for sensitive fungal isolates (mean EC\(50\) = 0.004 mg L\(^{-1}\); highest MIC = 0.03 mg L\(^{-1}\)).

For the second group of the 27 remaining orchards, for which the EC\(50\) values of the V. inaequalis spores ranged from 0.002 to 0.04 mg L\(^{-1}\) of kresoxim-methyl, the G143A mutation could be detected only in four of them.

### 3.3 Occurrence of the G143A mutation in French orchards from 2004 to 2007

From 2004 to 2007, infected leaves were sampled in 281 different commercial orchards in which fungicide treatments failed to limit the spread of apple scab, and 255 were successfully analysed by either PCR-based method. A total of 29 orchards were analysed over 2 years (22 orchards) or 3 years (7 orchards). In total, this represented 291 sample analyses. In 2004 and 2005 the samples were exclusively analysed using the CAPS method. In 2006, 67 samples were analysed by the CAPS method and 20 using AS-PCR (Table 4). Among the 67 initially analysed by CAPS, 21 were found to be positive and 46 to be negative for the mutation. Among these latter, 33 of them were reanalysed by AS-PCR, and five of them (15%) turned out to be positive for the mutation using this latter method. In 2007, all samples were analysed by AS-PCR.

#### Table 4. Prevalence of the Venturia inaequalis Cyt b G143A mutation in orchards sampled from 2004 to 2007, as revealed by either the CAPS method or allele-specific PCR (AS-PCR)

<table>
<thead>
<tr>
<th>Sampling year</th>
<th>No. of orchards</th>
<th>CAPS G143A mutation</th>
<th>AS-PCR G143A mutation</th>
</tr>
</thead>
<tbody>
<tr>
<td>2004</td>
<td>46</td>
<td>15</td>
<td>–</td>
</tr>
<tr>
<td>2005</td>
<td>125</td>
<td>43</td>
<td>–</td>
</tr>
<tr>
<td>2006</td>
<td>67</td>
<td>21</td>
<td>53(^a)</td>
</tr>
<tr>
<td>2007</td>
<td>–</td>
<td>–</td>
<td>33</td>
</tr>
<tr>
<td>2004–2007</td>
<td>238</td>
<td>79</td>
<td>86</td>
</tr>
</tbody>
</table>

\(^a\) Twenty unique samples \(\times\) 33 that were found to be negative in the CAPS assay.

\(^b\) Among which 5 were found to be negative by the CAPS method.

#### Table 3. Sensitivity to kresoxim-methyl of Venturia inaequalis spores collected in control orchards that had never been treated with strobilurins

<table>
<thead>
<tr>
<th>Orchard</th>
<th>Sampling year</th>
<th>Geographic origin(^a)</th>
<th>Germination(^b) (%)</th>
<th>EC(50)(^c) (mg L(^{-1}))</th>
<th>MIC(^d) (mg L(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>FR B</td>
<td>2005</td>
<td>Saint Marcel d’Urfé</td>
<td>10</td>
<td>0.006</td>
<td>0.03</td>
</tr>
<tr>
<td>FR B</td>
<td>2007</td>
<td>Saint Marcel d’Urfé</td>
<td>20</td>
<td>0.005</td>
<td>0.03</td>
</tr>
<tr>
<td>FR M</td>
<td>2005</td>
<td>Saint Marcel d’Urfé</td>
<td>12</td>
<td>0.002</td>
<td>0.01</td>
</tr>
<tr>
<td>FR M</td>
<td>2006</td>
<td>Saint Marcel d’Urfé</td>
<td>13</td>
<td>0.003</td>
<td>0.01</td>
</tr>
<tr>
<td>FR M</td>
<td>2007</td>
<td>Saint Marcel d’Urfé</td>
<td>14</td>
<td>0.006</td>
<td>0.03</td>
</tr>
<tr>
<td>FR V</td>
<td>2005</td>
<td>Saint Marcel d’Urfé</td>
<td>10</td>
<td>0.001</td>
<td>0.01</td>
</tr>
<tr>
<td>FR V</td>
<td>2007</td>
<td>Saint Marcel d’Urfé</td>
<td>15</td>
<td>0.005</td>
<td>0.03</td>
</tr>
<tr>
<td>VB</td>
<td>2004</td>
<td>Givors</td>
<td>28</td>
<td>0.004</td>
<td>0.03</td>
</tr>
<tr>
<td>CB</td>
<td>2004</td>
<td>Saint Jean le Vieux</td>
<td>20</td>
<td>0.005</td>
<td>0.01</td>
</tr>
</tbody>
</table>

| Mean (± SD) | 15.7 (±5.5) | 0.004 (±0.002) | 0.02 (±0.01) |

\(^a\) Localities are all located in the French administrative region Rhône-Alpes.

\(^b\) Percentage germination refers to the value obtained on fungicide-free medium.

\(^c\) EC\(50\) refers to the fungicide concentration that leads to a 50% inhibition of the germination rate.

\(^d\) MIC is the minimal concentration of fungicide that entirely inhibits spore germination.
from which samples were analysed: Midi-Pyrénées, Rhône-Alpes mutation was recorded in three (33%) regions out of the nine of the G143A mutation in multiple apple-producing regions in strobilurin treatments in these orchards. be detected over this time period, in spite of an alleged cessation for 12 of them. Except for one orchard, the mutation continued to 3 years, the mutation was detected in the first year of monitoring each of these four orchards. For the 29 orchards analysed over 2 or predominated among the different QoI resistance in the field have suggested that mutations conferring this phenotype could have been selected several times independently and/or were disseminating among pathogen populations.15 – 17

The present analyses clearly indicated the repeated occurrence of the G143A mutation in multiple apple-producing regions in France all through the sampling years (Fig. 7). Indeed, in 2004 the mutation was recorded in three (33%) regions out of the nine from which samples were analysed: Midi-Pyrénées, Rhône-Alpes and Provence-Alpes-Côte d’Azur. In 2005, five (42%) regions out of 12 contained the mutation, six (54%) out of 11 in 2006 and all seven in 2007. In the two administrative regions where the G143A mutation had prevailed since 2004 (the Midi-Pyrénées and Rhône-Alpes regions), a global increase in the percentages of orchards with the G143A mutation was noticed, ranging from 73% in 2004 to 86% in 2006 in Midi-Pyrénées and from 67% in 2004 to 84% in 2006 in Rhône-Alpes (Fig. 7).

4 DISCUSSION

As one of the major pathogens of apple trees, it is necessary to monitor the occurrence, origin and distribution of fungicide resistance in V. inaequalis in order to optimise the use of pesticides in apple tree orchards. Single-target-site QoI fungicides have become popular to control apple scab. Recent reports of QoI resistance in the field have suggested that mutations conferring this phenotype could have been selected several times independently and/or were disseminating among pathogen populations.15 – 17

As shown in this study, spor germination assays on fungicide-supplemented media, although effective to estimate resistance levels, fail to give interpretable results in many cases owing to very low spore viability in field-collected samples. Indeed, field samples have often been exposed to additional fungicides other than QoI molecules, which, in addition to delays between collection time in the field and processing in the laboratory, may affect spore viability. Nevertheless, spore germination assays demonstrate that, in about 25% of the samples that were processed, the failure properly to control apple scab could not be attributed to QoI resistance, as both the EC50 and the RF values for these samples were similar to the values recorded for samples collected in control orchards contaminated by QoI-sensitive V. inaequalis isolates.

Detection of the G143A mutation by the CAPS method and/or AS-PCR has already been reported for several plant pathogens such as Magnaporthe grisea (Hebert) Barr,18 Erysiphe graminis DC19 and Alternaria sp.,20 but not for V. inaequalis. The AS-PCR developed here proved to be at least 10 times more sensitive than the CAPS method based on the difference in detection level in the test samples containing different ratios of the mutated allele. This was confirmed in the samples from the orchards: the G143A mutation was detected in mixed lesion samples that were first interpreted as free of the mutation by CAPS. Nevertheless, this latter method may provide additional information compared with the AS-PCR, i.e. the presence of both the wild-type and the mutant Cyt b allele-bearing isolates occurring in a unique leaf sample. The digestion profiles indicate that in most cases (98%) the samples contain both the wild-type and the mutant Cyt b alleles. This situation may result from the coexistence in the samples of leaf lesions colonised by either mutant or wild-type sensitive strains. Alternatively, leaf lesions may have been colonised by heteroplasmic fungal strains containing in their mycelia a mixed population of mitochondria with or without the G143A mutation. Both hypotheses could be tested by analysing separately different lesions from the corresponding orchards as well as single spore isolates, as it cannot be ruled out that a single lesion could result from a coinfection by a QoI-sensitive and a resistant isolate. Heteroplasmy has been demonstrated for fungicide-resistant isolates originating from commercial orchards.15 

As anticipated, the present results clearly show a strong association between the occurrence of the G143A mutation and the detection of QoI resistance by in vitro spore germination
assays. Furthermore, this mutation was also observed in many other samples that could not be studied by the in vitro assay. Altogether, these results demonstrate that the main cause of the failure to control apple scab using QoI fungicides arises from the spread of fungal strains harbouring this mutation. In a few cases, discrepancies between the spore germination assays and the molecular detection of the G143A mutation were observed. A high level of resistance in the absence of the G143A mutation could indicate that an alternative resistance mechanism is operating. Other resistance mechanisms have indeed been reported in *V. inaequalis* and/or in other fungi. Change from phenylalanine to leucine at position 129 of the Cyt b polypeptide (mutation F129L) has been shown to occur in field populations of *Pyricularia grisea* Sacc.7,21 The present authors failed to detect this point mutation by sequencing the corresponding segment of the *V. inaequalis* gene amplified from one sample showing a moderate level of resistance without harbouring the G143A mutation (data not shown). The alternative oxidase pathway may also operate, thus leading to resistance, as previously demonstrated in *V. inaequalis*.10,14 although, in the present case, inclusion of SHAM in the spore germination medium did not modify strobilurin sensitivity (data not shown). Further experiments would be needed to identify the other mechanisms of QoI resistance in *V. inaequalis*.

In contrast, PCR-based detection of the G143A mutation in samples that were shown to be QoI-sensitive in spore germination assays is more difficult to interpret. It could be hypothesised that these samples were a mixture of lesions colonised by predominantly sensitive isolates versus QoI-resistant ones, and that only the former contributed, owing to germination bias, to the spore germination assay, whereas DNA from both lesions contributed to the molecular assay.

In addition to being more reliable than the spore germination assays, molecular monitoring is less time consuming. It was performed on 291 samples collected in the main apple-producing regions of France in orchards in which apple scab could not be controlled by the use of strobilurin. These orchards are, however, not representative of French orchards in terms of apple scab control. Multiple detection of QoI resistance in the French apple-producing regions raises the epidemiological question about its origin, i.e. frequent occurrence of independent G143A mutational events versus the long-distance dissemination of a single or of very few G143A alleles.

**SUPPORTING INFORMATION**

Supporting information may be found in the online version of this article.

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