



HAL
open science

**Lack of experimental evidence to support mcr-1-
positive escherichia coli strain selection during oral
administration of colistin at recommended and higher
dose given by gavage in weaned piglets**

Alexis Viel, Jérôme Henri, Agnès Perrin-Guyomard, Julian Laroche, William
Couet, Nicolas Grégoire, Michel Laurentie

► **To cite this version:**

Alexis Viel, Jérôme Henri, Agnès Perrin-Guyomard, Julian Laroche, William Couet, et al.. Lack of experimental evidence to support mcr-1- positive escherichia coli strain selection during oral administration of colistin at recommended and higher dose given by gavage in weaned piglets. *International Journal of Antimicrobial Agents*, 2017, 51 (1), pp.128-131. 10.1016/j.ijantimicag.2017.04.013 . anses-01570163

HAL Id: anses-01570163

<https://anses.hal.science/anses-01570163>

Submitted on 22 Jun 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Lack of experimental evidence to support *mcr-1*-positive *Escherichia coli***
2 **strain selection during oral administration of colistin at recommended and**
3 **higher dose given by gavage in weaned piglets**

4
5 Alexis Viel^{a,b,c}, Jérôme Henri^c, Agnès Perrin-Guyomard^c, Julian Laroche^{a,d}, William
6 Couet^{a,b,d}, Nicolas Grégoire^{a,b,*}, Michel Laurentie^c

7
8
9 ^aInserm U1070, Pôle Biologie Santé - Bât. B36/37, 1 rue Georges Bonnet, Poitiers, France

10 ^bUniversité de Poitiers, UFR Médecine-Pharmacie, 6 rue de la Milétrie, Poitiers, France

11 ^cAnses, Laboratoire de Fougères, 10B Rue Claude Bourgelat, Fougères, France

12 ^dCHU Poitiers, 2 rue de la Milétrie, Poitiers, France

13

14

15

16

17

18

19 *** Corresponding author**

20 Tel.: +33 5 49 36 64 36.

21 E-mail address: nicolas.gregoire@univ-poitiers.fr

22

23

24

25 **Abstract**

26 In this study, we assessed the selective effect of colistin orally administered to healthy weaned
27 piglets harbouring an intestinal *mcr-1*-positive *Escherichia coli* strain. Maximum
28 recommended dose and a higher dose often used in European pig farms were given by
29 gavage. No selection of the *mcr-1*-positive strain was observed in our controlled conditions
30 whatever the dose. Further investigations in real farming conditions seem necessary.

31

32 **Keywords:** colistin; *mcr-1*; *Escherichia coli*; piglets; selection

33

34 **1. Introduction**

35 Colistin is an old polypeptidic antibiotic widely used in food-producing animals, especially in
36 pig production as oral group treatment and metaphylaxis against Enterobacteriaceae digestive
37 infections after weaning. Colistin is also used in human medicine as a last resort antibiotic
38 against multi-drug bacteria. The first plasmid-mediated colistin resistance gene (*mcr-1*)
39 discovered in China at the end of 2015 [1] has raised concern about the risk of spread of this
40 resistance. Few months after, *mcr-1* was detected in all continents, both in human and animals
41 [2]. European Medicines Agency emphasized the need of reducing colistin use in animal and
42 proposed to class colistin as critically important antimicrobials [3]. Scientific community
43 lacks of in vivo data about *mcr-1*, especially in commensal Enterobacteriaceae to properly
44 characterize the public health risk. We assessed here in piglets the selective effect of
45 controlled colistin oral treatments on commensal intestinal *Escherichia coli* harbouring a *mcr-*
46 *1*-positive strain with monitoring of faecal concentrations.

47

48 2. Materials and methods

49 2.1. Animals and housing

50 Fifteen Large White-Landrace-Piétrain piglets were used to carry out the experiment, with no
51 history of antimicrobials treatments. They were weaned at 21 days old and then fed with a
52 standard non-medicated ration and had free access to water. After 5 days of collective
53 housing, they were put in individual boxes with no possible contact (8 days before treatment,
54 D-8).

55

56 2.2. Bacterial strain and inoculation

57 The original strain was a colistin-resistant *E. coli* (MIC = 8 mg/L) isolated from the intestines
58 of a healthy pig sampled in a French slaughterhouse [4]. This strain harbouring *mcr-1*
59 (confirmed by PCR) was made rifampicin-resistant (MIC > 512 mg/L) by spontaneous
60 mutation before inoculation and named ECmcr1+. The inoculation phase consisted of three
61 gavages (at D-7, D-5 and D-2) of 5 mL of about 10^7 CFU/mL of ECmcr1+ in saline
62 suspension.

63

64 2.3. Experimental treatment and sampling

65 Piglets were randomly divided into three groups of 5 animals and force-fed with colistin
66 sulphate (Acti-coli, Biové, Arques, France) using a polyethylene tube, from D0 to D4 (5
67 days). RD group (for maximum Recommended Dosage) received 100 000 UI/kg/day [5]
68 equivalent to 3 mg/kg/day of colistin base activity (CBA) [6], given twice a day as 1.5 mg/kg
69 in 5 mL solution; HD group (for Higher Dosage often found in pig farms [7]) received
70 200 000 UI/kg/day equivalent to 6 mg/kg/day of CBA, given twice a day as 3 mg/kg in 5 mL
71 solution; the placebo group received water. Fresh faecal samples were taken on mornings
72 (after anal stimulation) from the day before ECmcr1+ inoculation (D-8) until 19 days after the

73 end of colistin treatment (D23). This experiment was approved by the ComEth
74 Anses/ENVA/UPEC n°16 (French ethical committee) under the reference APAFIS#2905-
75 2015112717486085.

76

77 *2.4. Microbiological analysis and colistin assay*

78 About 1 g of each fresh faecal sample was diluted in saline solution. Selected dilutions were
79 plated on Mac Conkey agar (BD, Le Pont de Claix, France) alone or supplemented with 200
80 mg/L of rifampicin (Sigma, Saint-Quentin Fallavier, France) in order to count total *E. coli* and
81 ECmcr1+, respectively.

82

83 Another specimen (1 g of faeces) from D0 to D9 was kept at -20°C until colistin was assayed
84 by a LC-MS/MS method adapted from previous works [8, 9]. Briefly, faeces were mixed with
85 blank plasma and diluted in 10 mL of acetonitrile with 6% of trichloroacetic acid. After
86 vortexing, centrifugation and evaporation, dry matter was diluted in buffer (pH=7.2) with
87 blank plasma and loaded on Oasis HLB Catridges (Waters, Milford, MA, USA). After
88 washing and eluates evaporation, residues were analysed by HPLC-MS/MS with a limit of
89 quantification (LOQ) of 1 µg/g of faeces. Quality controls were prepared at 2.5, 12.5 and 18.8
90 µg/g of faeces.

91

92 *2.5. Statistical analysis*

93 Mean values of colistin faecal concentrations of the two treated groups were compared using
94 a Student-T test. Mean values of total faecal *E. coli* and of ECmcr1+ for treated groups were
95 compared to those of the placebo group using a Student-T test. All statistical analyses were
96 carried out using R 3.3.2 [10].

97

98 **3. Results**

99 High faecal colistin concentrations were measured with mean values greater than or equal to
100 about 200 µg/g of faeces from D2 to D5 in the two treated-groups (Table 1). No significant
101 differences were observed between these groups (T-test) due to the high inter-individual
102 variability. Three days after the end of treatment (D7), faecal concentrations were reduced by
103 about 100-fold and fell below the LOQ after 2 days more (D9).

104

105 No significant differences were found for total faecal *E. coli* (EC_{tot}) population between
106 groups before colistin administration (from D-8 to D0, Fig. 1A). We observed a slight but
107 non-significant decrease of EC_{tot} during the treatment phase for the placebo group.

108 Conversely, a stronger reduction of EC_{tot} was noticed for each colistin-treated group between
109 D0 and D3 but only significant for HD group (compared to placebo, Fig. 1A). Then, a return
110 to initial level of EC_{tot} was observed 2 and 3 days after last colistin dose (D4) for RD and HD
111 group, respectively.

112

113 After the inoculation phase (D-7 to D-2), EC_{mcr1+} reached up to 2 % of EC_{tot} (Fig. 1B).
114 However 7 of 15 piglets had EC_{mcr1+} count lower to the LOQ (1.2 log CFU/g of faeces) just
115 before treatment ; for 5 of them the strain was undetectable during all the experiment (2 in HD
116 group, 2 in placebo group and 1 in RD group). Mean EC_{mcr1+} levels were equivalent
117 between the three groups before the first colistin administration (D0). Overall, EC_{mcr1+} level
118 compared favourably between each group i.e. remained relatively constant during and after
119 treatment.

120

121 4. Discussion

122 To our knowledge, this is the first in vivo study exploring the selective effect of colistin on
123 *mcr-1*-positive *E. coli* in pigs. Previous studies already attested a rare emergence of colistin-
124 resistant *E. coli* after oral colistin treatment of healthy [7] and sick piglets [11], therefore we
125 neglected it. Exogenous resistant bacteria inoculation in treated pigs already supported the
126 selective effect of other antimicrobials in these animals [12]. In order to control the colistin
127 doses, we chose to force-feed the piglets for treatment. The absence of significant differences
128 of faecal colistin concentrations between HD and RD groups was likely due to measurement
129 uncertainty. However, these results seemed relevant: with a daily faecal excretion of piglets of
130 around 250 g [13], we can estimate that about 60 and 95 % of the colistin dose was recovered
131 in faeces for HD and RD group. This is consistent with the poor absorption of colistin
132 sulphate after oral treatments in pigs [5]. In addition, we chose to induce a rifampicin
133 resistance in ECmcr1+ in order to monitor precisely this strain and due to the poor accuracy
134 of colistin-supplemented media for resistant Enterobacteriaceae isolation [7, 11].

135

136 The initial decrease of EC_{tot} level before treatment and the slight continuous one of placebo
137 group were probably due to the weaning process that disturbed the microbiota equilibrium
138 [14]. The high colistin treatment induced a significant reduction of EC_{tot} counts compared to
139 the placebo group (Fig. 1A), but the recommended dose poorly affected it. Considering the
140 high faecal concentrations during the treatment phase (> 200 µg/g), greater reduction of *E.*
141 *coli* would be expected. Colistin high adsorption to faecal fibres could lead to a dramatic
142 decrease of its effect [15]. In addition, for polymyxin B, up to 90 % of the initial dose could
143 be reversibly bound to faeces [16]. Although this was shown with a bioassay method and not
144 with colistin, the real active colistin concentrations in our experiment should be greater than
145 20 µg/g of faeces. This is normally still enough to reduce EC_{tot} population as it is composed

146 of majority of colistin-sensitive strains ($CMI \leq 2$ mg/L). Therefore, the absence of significant
147 difference of the EC_{tot} evolution between the two treated groups is still unclear. The small
148 number of animal per group is perhaps a limiting factor. Moreover, inter and intra-individual
149 variabilities of faecal colistin concentrations were high, probably due to a heterogeneous
150 colistin distribution within faeces. Therefore, the actual effective concentrations of colistin, to
151 which bacteria were exposed within the digestive tract, are mostly unknown.

152

153 In the placebo group, the counts of EC_{mcr1+} were low but remained stable enough over the
154 experiment (Fig. 1B). No selective effect of colistin in treated piglets was observed as
155 EC_{mcr1+} stayed at a constant level over time, whatever the dose. This suggests that, in
156 standardized conditions, bacteria were exposed during a too short period to concentrations
157 within the selection window, i.e. between the MICs of indigenous *E. coli* and of EC_{mcr1+}
158 (8 mg/L). In comparison, when colistin is administered on a large scale through water or feed
159 on pig farms, colistin concentration in faeces are lower [7] and much more variable [17];
160 therefore the probability to select strains harbouring *mcr-1* increases. Despite this, no real
161 outbreak of *mcr-1* positive strains exists in pig production in Europe [2] where a low
162 prevalence of *mcr-1* positive *E. coli* is found (e.g. 0.5 % in French pig farms) [4]. In contrast,
163 a high prevalence is observed in Asia (more than 20 % in China) [2], where colistin has been
164 used for decades as growth-promoter for piglets [18]. This sub-therapeutic dose (about 4-5
165 times lower than therapeutic use [19]) administered during a long period is likely to give
166 digestive concentrations reaching the selection window of *mcr-1* positive strains. Awareness
167 of this high risk led China to ban colistin as promoter very recently [20]. This hypothesis
168 should deserve further considerations.

169

170

171 **5. Conclusions**

172 In conclusion, under this experimental setting in piglets, oral dosing with colistin did not
173 induce selection of *mcr-1* positive strains. Further investigations should be necessary to
174 confirm this observation in farming conditions (with colistin given via food or water and
175 piglets having contact between them). Meanwhile, a responsible and careful use of colistin is
176 required to preserve this last-resort antimicrobial agent.

177

178 **Acknowledgments**

179 The authors would like to thank Jean-Guy Rolland and Mireille Bruneau, Anne De Courville,
180 Karine Deleurme, Pamela Houée, Catherine Poirier for their technical assistance.

181

182 **Declarations**

183 **Funding:** Alexis Viel was supported by a doctoral fellowship from the French National
184 Institute of Health and Medical Research (Inserm) and the French Agency for Food,
185 Environmental and Occupational Health & Safety (Anses).

186 **Competing Interests:** None

187 **Ethical Approval:** This experiment was approved by the ComEth Anses/ENVA/UPEC n°16
188 (French ethical committee) under the reference APAFIS#2905-2015112717486085

189

190

191

192 **References**

- 193 [1] Liu Y-Y, Wang Y, Walsh TR, Yi L-X, Zhang R, Spencer J, et al. Emergence of plasmid-
194 mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a
195 microbiological and molecular biological study. *The Lancet infectious diseases*. 2016;16:161-
196 8.
- 197 [2] Skov R, Monnet D. Plasmid-mediated colistin resistance (*mcr-1* gene): three months later,
198 the story unfolds. *Euro surveillance : bulletin Europeen sur les maladies transmissibles =*
199 *European communicable disease bulletin*. 2016;21:30155.
- 200 [3] Updated advice on the use of colistin products in animals within the European Union:
201 development of resistance and possible impact on human and animal health.: European
202 Medicines Agency (EMA); 2016.
- 203 [4] Perrin-Guyomard A, Bruneau M, Houée P, Deleurme K, Legrandois P, Poirier C, et al.
204 Prevalence of *mcr-1* in commensal *Escherichia coli* from French livestock, 2007 to 2014.
205 *Euro surveillance : bulletin Europeen sur les maladies transmissibles = European*
206 *communicable disease bulletin*. 2016;21.
- 207 [5] Guyonnet J, Manco B, Baduel L, Kaltsatos V, Aliabadi M, Lees P. Determination of a
208 dosage regimen of colistin by pharmacokinetic/pharmacodynamic integration and modeling
209 for treatment of GIT disease in pigs. *Research in veterinary science*. 2010;88:307-14.
- 210 [6] Nation RL, Li J, Cars O, Couet W, Dudley MN, Kaye KS, et al. Consistent Global
211 Approach on Reporting of Colistin Doses to Promote Safe and Effective Use. *Clinical*
212 *Infectious Diseases*. 2014;58:139-41.
- 213 [7] Fleury MA, Jouy E, Eono F, Cariolet R, Couet W, Gobin P, et al. Impact of two different
214 colistin dosing strategies on healthy piglet fecal microbiota. *Research in Veterinary Science*.
215 2016;107:152-60.

216 [8] Van den Meersche T, Van Pamel E, Van Poucke C, Herman L, Heyndrickx M, Rasschaert
217 G, et al. Development, validation and application of an ultra high performance liquid
218 chromatographic-tandem mass spectrometric method for the simultaneous detection and
219 quantification of five different classes of veterinary antibiotics in swine manure. *Journal of*
220 *Chromatography A*. 2016;1429:248-57.

221 [9] Gobin P, Lemaitre F, Marchand S, Couet W, Olivier JC. Assay of colistin and colistin
222 methanesulfonate in plasma and urine by liquid chromatography-tandem mass spectrometry.
223 *Antimicrobial agents and chemotherapy*. 2010;54:1941.

224 [10] R Core Team. R: A language and environment for statistical computing. R Foundation
225 for Statistical Computing, Vienna, Austria; 2016.

226 [11] Rhouma M, Beaudry F, Thériault W, Bergeron N, Beauchamp G, Laurent-Lewandowski
227 S, et al. In vivo therapeutic efficacy and pharmacokinetics of colistin sulfate in an
228 experimental model of enterotoxigenic *Escherichia coli* infection in weaned pigs. *Veterinary*
229 *Research*. 2016;47:58.

230 [12] Cavaco LM, Abatih E, Aarestrup FM, Guardabassi L. Selection and Persistence of CTX-
231 M-Producing *Escherichia coli* in the Intestinal Flora of Pigs Treated with Amoxicillin,
232 Cefotiofur, or Cefquinome. *Antimicrobial Agents and Chemotherapy*. 2008;52:3612-6.

233 [13] Pouliot F, Godbout S, Dufour V, Vob Bernuth R, Hill J. Évaluation de l'efficacité d'un
234 système de séparation fèces-urine sous caillebotis en engraissement: bilan de masse et
235 caractérisation des sous-produits. *Journées Rech Porcine*. 2005;37:45-50.

236 [14] Swords WE, Wu C-C, Champlin FR, Buddington RK. Postnatal changes in selected
237 bacterial groups of the pig colonic microflora. *Neonatology*. 1993;63:191-200.

238 [15] Van Saene JJ, Van Saene HK, Stoutenbeek CP, Lerk CF. Influence of faeces on the
239 activity of antimicrobial agents used for decontamination of the alimentary canal.
240 *Scandinavian journal of infectious diseases*. 1985;17:295-300.

241 [16] Hazenberg M, Pennock-Schröder A, Van de Merwe J. Reversible binding of polymyxin
242 B and neomycin to the solid part of faeces. *Journal of Antimicrobial Chemotherapy*.
243 1986;17:333-9.

244 [17] Soraci AL, Amanto F, Tapia MO, de la Torre E, Toutain P-L. Exposure variability of
245 fosfomycin administered to pigs in food or water: impact of social rank. *Research in*
246 *veterinary science*. 2014;96:153-9.

247 [18] Kim DP, Saegerman C, Douny C, Dinh TV, Xuan BH, Vu BD, et al. First survey on the
248 use of antibiotics in pig and poultry production in the Red River Delta region of Vietnam.
249 *Food and Public Health*. 2013;3:247-56.

250 [19] Rhouma M, Beaudry F, Letellier A. Resistance to colistin: what is the fate for this
251 antibiotic in pig production? *International journal of antimicrobial agents*. 2016;48:119-26.

252 [20] Walsh TR, Wu Y. China bans colistin as a feed additive for animals. *The Lancet*
253 *Infectious Diseases*. 2016;16:1102-3.

254

255 **Figure Legend**

256 **Fig 1:** Mean (\pm SD) of counts of total faecal E.coli (A) and ECmcr1+ (B) before, during and
257 after oral colistin (or water) administrations for HD group (green), RD group (blue) and
258 placebo group (red). Vertical arrows indicate ECmcr1+ inoculations. The vertical dotted lines
259 indicate the treatment period (D0 to D4). Horizontal dashed line represents the limit of
260 quantification (log 1.2) and data below this value were arbitrary put to value log 0.6. Similar
261 profiles were obtained when data below LOQ were fixed at zero or at LOQ. Significant
262 differences from placebo group with Student test: * ($p < 0.05$); ** ($p < 0.01$)
263

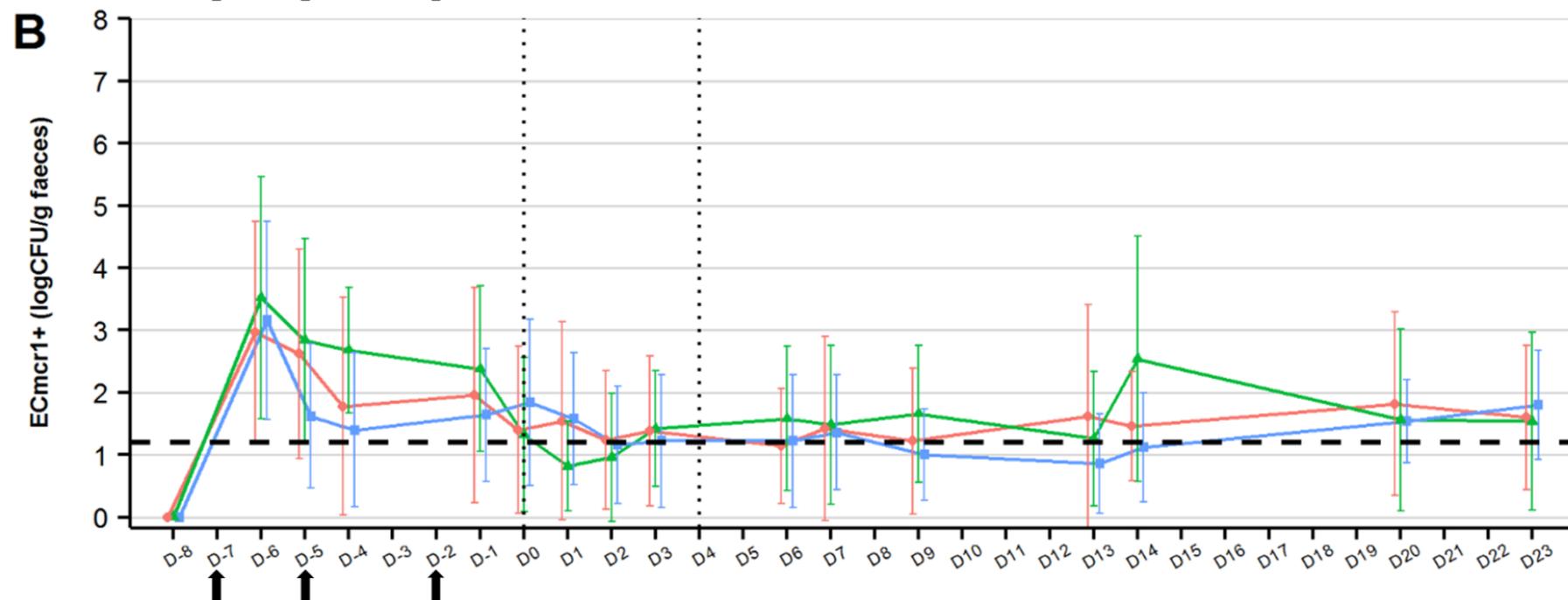
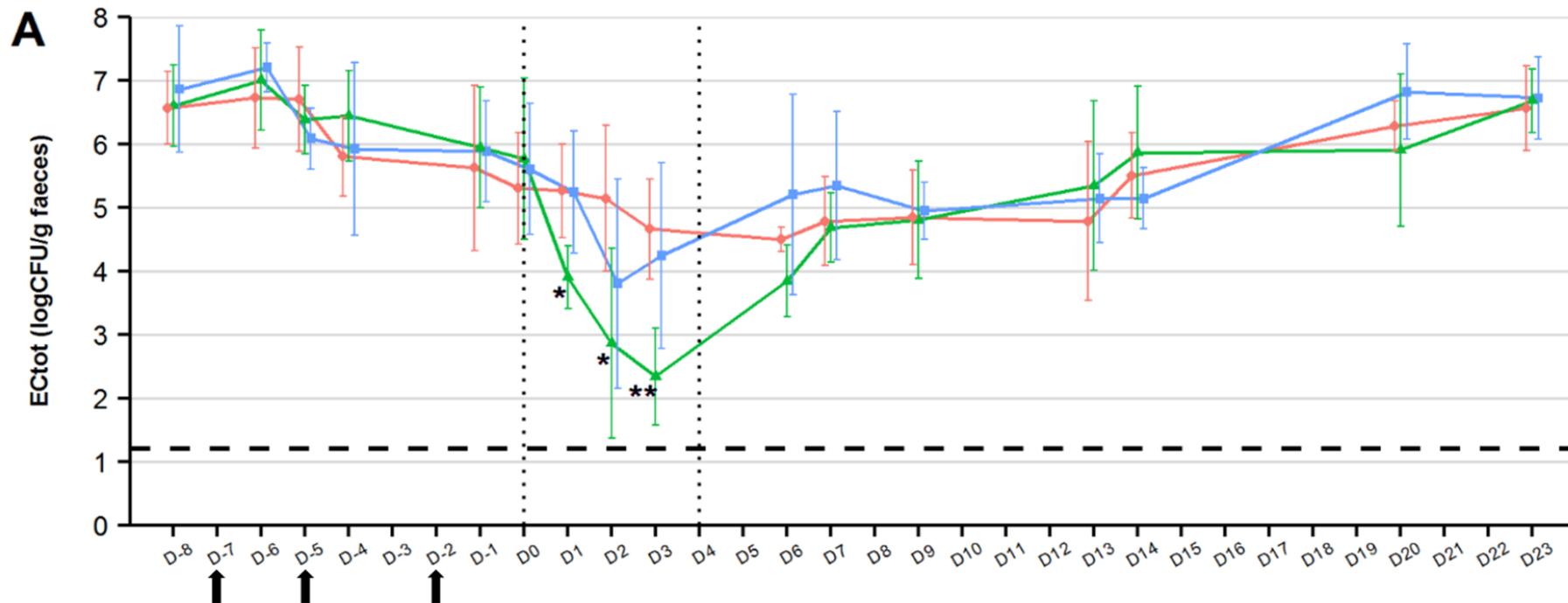


TABLE 1: Faecal colistin concentrations over time in each treated-group^a.

Treatment Group	Mean \pm SD (and range) in μg of colistin base/g of faeces per day.									
	D0	D1	D2	D3	D4	D5	D6	D7	D9	
Recommended Dose (100,000 UI/kg/day)	0	93.2 \pm 56.0 (28.1 - 149.9)	193.6 \pm 144.5 (75.5 - 381.5)	217.4 \pm 103.3 (74.2 - 327.0)	197.1 \pm 54.7 (128.9 - 261.2)	262.5 \pm 57.8 (212.4 - 342.1)	42.3 \pm 22.8 (14.9 - 72.0)	1.8 \pm 0.3 (1.6 - 2.2)	< LOQ ^b	
High Dose (200,000 UI/kg/day)	0	119.6 \pm 55.2 (61.2 - 188.9)	267.9 \pm 78.3 (169.7 - 354.6)	268.9 \pm 119.2 (155.0 - 470.9)	373.4 \pm 190.2 (141.6 - 669.5)	249.7 \pm 109.1 (106.3 - 372.4)	80.3 \pm 103.8 (15.0 - 264.8)	3.4 \pm 1.6 (1.7 - 5.7)	< LOQ ^b	

^a Placebo group is not mentioned as all concentrations were null^b LOQ: 1 μg /g of faeces